GLOBAL ANALYSIS OF NON-SYNONYMOUS SNPs IN MONKEYPOX VIRUS

Oluwaseyi Abraham Olawale, Oguntomi Ayomide Samuel, Zaimat Sanni, Precious Oyeboade, Shabana Bibi, Maryam Idrees, Meenakshi Agarwal

Oluwaseyi Abraham Olawale, Gen'Omics Research Hub (GENOMAC HUB), Ogbomoso, Nigeria Oguntomi Ayomide Samuel, Gen'Omics Research Hub (GENOMAC HUB), Ogbomoso, Nigeria Zaimat Sanni, Department of Medical Laboratory Science, Ladoke Akintola University of Technology, Ogbomoso, Nigeria. Precious Oyeboade, Department of Physiology, Ladoke Akintola University of Technology, Ogbomoso, Nigeria. Shabana Bibi, Department of Biosciences, Shifa Tameer-e-Millat University, Islamabad 44000, Pakistan Maryam Idrees, Department of Microbiology, Quaid-i-Azam University, Islamabad, Pakistan. Meenakshi Agarwal, Florida A&M Universit, Tallahassee, Fl, USA. Correspondence Author: Oluwaseyi Olawale.

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ABSTRACT

In May of 2022, it was determined that a virus known as monkeypox (MPX) had spread over the world. The MPXV strain responsible for 87000 illnesses and 112 fatalities has been identified. This strain had a global average of 38.35 mutations per virus sample, with the 2022 epidemic seeing the highest MPXV mutation rate. Our aim was to use publicly accessible NCBI (National Center for Biotechnology Information) data to explore the genetic diversity and evolutionary history of monkeypox viruses (MPXVs).

In-depth comparisons of the genomic characteristics of 376 publicly available complete MPXV genome sequences were performed. Nucleotide sequence variation, gene order, mapping, alignment, and mapping accuracy were among these features. A large amount of sequence conservation was discovered in the MPXV genomes by the meta-analysis, with only a few minor changes between isolates. Multiple genomic regions were discovered to be highly conserved among all MPXV strains, suggesting an essential function in viral replication and persistence. The use of phylogenetic analyses to trace the evolutionary relationships of MPXV strains revealed that the virus has diverged into several subgroups with distinct genetic makeups.

Conclusion: Our findings showed that non-synonymous single nucleotide polymorphisms may be impacting the evolution of MPXV isolates that are dispersed globally and promoting improved adaptability and human-to-human transmissibility.

1. INTRODUCTION

Poxviridae viruses are oval-shaped viruses with large double-stranded DNA genomes and the ability to replicate in the cytoplasm of host cells rather than the nucleus (1). Poxvirus infections typically result in skin nodules, lesions, or a widespread rash (2). The orthopoxvirus genus includes closely related poxviruses such as monkeypox virus (MPXV), variola virus (VARV), vaccinia virus (VACV), and cowpox virus (CPV) (3). As a result, MPXV is defined as a zoonotic orthopox DNA virus that is the causative agent of monkeypox illness (MPX), which was first described in the Democratic Republic of the Congo (DRC) (previously known as Zaire) in 1970 (5).

MPX causes a variety of signs and symptoms, ranging from dermatologic to systemic issues. Fever is usually the initial sign of infection, followed by the growth of many papular, vesiculopustular, and ulcerative lesions on the face and body, as well as a detectable lymphadenopathy. Encephalitis, pneumonitis, keratitis, and subsequent bacterial infections are the most prevalent problems linked with MPX (6, 7). Close contact with an infected animal (rodents are considered to be the principal animal reservoir for human transmission), large respiratory droplets, close or direct touch with skin lesions, or possibly contaminated fomites can all result in MPXV transmission (8). Because orthopoxviruses are serologically cross-reactive, polymerase chain reaction (PCR) is favored over antigen and antibody detection methods. The European Medicines Agency (EMA) approved tecovirimat, a smallpox (VARV) antiviral drug, for the treatment of MPX in 2022, although it is not yet widely available. Smallpox immunizations provide 85% protection against MPXV, and previously vaccinated persons may experience milder symptoms. An improved two-dose vaccination based on vaccinia virus was approved for the prevention of MPX in 2019, but it is not yet widely available (6).

MPXV is becoming more prevalent in the human population, which could lead to genetic adaptation for effective transmissibility and pathogenicity (5). From January 1 to June 22, 2022, 3413 laboratories confirmed MPXV cases and one fatality were reported from five WHO Regions comprising 50 countries/territories. WHO The European Region had the highest number of laboratory confirmed cases (86%; 2933/3413), followed by the Americas Region (11%; 381/3413), Africa (2%; 73/3413), the Eastern Mediterranean Region (1%; 15/3413), and the Western Pacific Region (1%; 11/3413).In Nigeria, one fatality was recorded in the second quarter of 2022 (9).

Several variables have contributed to the success of the MPXV 2022 outbreak. The successful eradication of VARV (smallpox virus) under the auspices of WHO resulted in the discontinuation of VARV vaccine, resulting in an increase in the population vulnerable to VARV and other phylogenetically related orthopoxviruses (10). In addition to the immunological and ecological niche vacated by VARV (11), suspected MPXV genome variations, relaxation in preventive measures against corona virus disease 2019 (covid-2019), restoration of international travel, and sexual interactions associated with large gatherings (12) may have aided MPXV reemergence and spread. Thornhill et al. investigated a case series of MPXV infection across 5 continents and 16 countries from April to June 2022 and observed that 98% of the infected people were gay or bisexual men, 75% were Caucasians, and 41% had an accompanying human immunodeficiency virus (HIV) infection. The average age of those afflicted was 38 years (13).

MPXV is genetically divided into two clades (14): West Africa (WA) and Congo Basin (CB); whole genome study found that the WA and CB lineages are separated by a 900 bp difference. MPXV has been shown to gain or lose genetic material in studies (11); nevertheless, based on the stability of the double-stranded DNA genome, MPXV should have a low mutation rate (15). Wherever possible, MPXV DNA from the present outbreak is being sequenced; preliminary data from polymerase chain reaction (PCR) assays of MPXV isolates from the current outbreak suggest the presence of genes associated with the WA clade (9). For the first time, the spread of MPXV has spread outside of West and Central Africa; it has been proposed that the successive loss of genetic factors unimportant for human pathogenesis served as a mechanism that resulted in the emergence of highly adapted MPXV with increased virulence and the ability to transmit efficiently and rapidly from person to person. As a result, the genetic diversity between the freshly sequenced MPXV isolates and the Zaire-96-I-6 strain is reported in this paper. The goal of this study is to investigate potential variants, anticipate impacts, and conduct phylogenetic analysis.

2. MATERIALS AND METHODS:

2.1 WORKFLOW



2.2 DATA

The SNP analysis was performed using whole genome sequences of MPXV isolates. WGS helps to detect copy number variations, rearrangement and structural variants. For variant analysis, Whole genome sequences of 376 MPXV isolates that were submitted to the NCBI (National Center for Biotechnology Information) from May to July 2022 were retrieved [16].

2.3 SNPs Analysis, Mapping and Alignment

In this study, SNPs analysis was carried out on the mutation of Monkeypox virus in relation to the current Global Pandemic of Monkeypox virus. Using Clade 1 as isolate which is monkeypox virus gotten from Zaire-96-I-16[15]. Whole genome sequences were mapped against Zaire-96-I-16 complete genome as reference sequence using geneious mapper v9.0.5 because it produces superior results and can correctly align structural variants [17], with medium sensitivity, specifying the time it takes to assemble and the accuracy of the assembly to create contig. Protein coding prediction was carried out using EMBOSS 6.5.7, tool, tcode on geneious software v9.0.5

2.4 SNPs Calling and Effect Prediction

SNPs are possible changes that, occurs between the sample and reference sequence. Using the Geneious Software v9.0.5 [17], we imported the downloaded WGS Sequences and conducted the alignment of 376 sequences using clade 1 as the reference (Zaire-96-I-16). The variant

calling generated for all samples used the geneious mapper v9.0.5 with a Minimum variant frequency of 0.6 and a Maximum Variant P-value of 10,000,000,000, a Minimum Strand-Bias P-value of 0.000001, when exceeding 65% bias value to determine the certainty of each association to the reference sequence.

2.5 PHYLOGENETIC ANALYSIS

Study of geographical pattern to determine the pattern of mutation and the common predecessor of each strain of all 2022 monkeypox virus strains reveals various genetic profiles and that random mutations occurred during the evolution process. Active surveillance has obtained monkeypox disease prevalence and topographical reach but there is no available information about virus diversity. Comparative analysis of strains was performed on NCBI based on statistical analysis to determine positions of important differentiation between the strains. We retrieved 376 whole genome sequences of humans from May to July of 2022 on the National Center of Biotechnology Information (NCBI) and edited using Interactive Tree Of Life (ITOL) We obtained 13 clades to discover the common predecessor of each strain of all 2022 monkeypox virus strains.

3 RESULTS

3.1 GENES

According to data on the monkeypox virus, the B17R mutation has been shown to occur multiple times, while the N4R, B11R, J2L, F8L, and B18R mutations have each been found to occur only once. Mutations point to the virus's advanced evolutionary state and host adhesion. J2L, B17R, B18R, B11R, F8L, A27L, and N4R are only some of the genes that can be altered. There are seven separate instances of variance in the B17R gene, but only one instance in each of the other six genes. that's a total of 13 distinct permutations. None of the 33 sequences had the same length. There are 27 one-sequences, 3 two-sequences, and 2 four-sequences. The CDS opinions diverge. The CDS locations of 20 sequences are absent, while those of the remaining 13 sequences range widely. This study aimed to determine the structural and functional impact of missense mutations on the N4R gene.

3.2 SNPs Analysis

On the basis of result similarity, all sequences are 400,483 in length, with an elevated standard of resemblance of 71.4% pair-wise after map to reference. SNPs calling was conducted using the geneious software v9.0.5. While 33 SNPs were obtained; however, 26 SNPs which were synonymous (not causing a change in amino acid) were not considered as it was assumed that synonymous SNPs were not consequential because the primary sequence of the protein is preserved. SNPs were allocated identification based on their positions in the genome of the reference strain. For instance, SNP which locates at position 4567 is named after SNP-4567. Ultimately, each variant at the interval of 1 is separated on the premise of the different positions of sequence nucleotide polymorphism (Transversion, Transition), genes and the effect on protein structure. Variant analysis of MPXV isolates revealed missense mutations; a missense mutation involves a point mutation in which a single nucleotide change results in a codon that encodes a different amino acid, as shown in Figure 1a and 1b.

3.3 Identification of nsSNPs on a Coding Region of Protein

We utilized the geneious software to conduct joint SNPs analysis to explore variants. The SNPs which have no effect on amino acid were filtered out and 7 non synonymous SNPs (alteration of amino acid) were found in the coding region. The nonsynonymous single nucleotide polymorphisms hereby referred to as nsSNPs includes, SNP191565, SNP2723, SNP175877, SNP169783, SNP54359, SNP176386, SNP177993. The nsSNPs resulted in amino acid substitution at the coding region.

3.4 SNPs With Effect on Protein Structure

SNP191565, with variant A as reference nucleotide and G as the variant nucleotide (Adenine-Guanine) encodes a codon change (ACA-GCA) resulting to a substitution of the amino acid from Threonine(T) to Alanine(A), with protein id (NP_536615.1), a length of 1 and locus tag (MPXVgp188) at a variant frequency of 61.2% and a variant P-Value 1.6E-47. SNP2723, with variant A as reference nucleotide and T as the variant nucleotide (Adenine-Thymine) encodes a codon change (TTA-ATA) resulting to a substitution of the amino acid from Leucine(L) to Isoleucine(I), with protein id (NP_536429.1), a length of 1 and locus tag (MPXVgp002) at a variant frequency of 75.5% and a variant P-Value 6.9E-61. SNP175877, with variant G, as

reference nucleotide and A as the variant nucleotide (Guanine-Adenine) encodes a codon change (GGA-GAA) which results to a substitution of the amino acid from Glycine(G) to Glutamate(E), with protein id (NP_536605.1), a length of 1 and locus tag (MPXVgp178) at a variant frequency of 61.1% and a variant P-Value 4.2E-52. SNP169783, with variant G as reference nucleotide and A as the variant nucleotide (G-A) encodes a codon change (GAA-AAA) resulting to substitution of the amino acid from Glutamate(E) to Lysine (K), with protein id (NP_536599.1), a length of 1 and locus tag (MPXVgp172) at a variant frequency of 79.6% and a variant P-Value 8.6E-76. SNP54359, with variant C as reference nucleotide and T as the variant nucleotide (Cytosine-Thymine) encodes a codon change (GAT-AAT) resulting to a substitution of the amino acid from Aspartate(D) to Asparagine(N), with protein id (NP_536484.1), a length of 1 and locus tag (MPXVgp057) at a variant frequency of 84.5% and a variant P-Value 9.7E89. SNP176386, with a variant G, as reference nucleotide and A as variant nucleotide (Guanine-Adenine) encodes a codon change (GAC-AAC) resulting to a substitution of the amino acid from Aspartate (D) to Asparagine (N), with protein id (NP_536605.1), a length of 1 and locus tag (MPXVgp178) at a variant frequency of 61.1% and a variant P-Value 4.2E-52. SNP177993, with a variant C as reference nucleotide and T as variant nucleotide (Cytosine-Thymine) encodes a codon change (GCA-GTA) which results to a substitution of amino acid from Alanine (A) to Valine (V), with protein id (NP_536606.1), a length of 1 and locus tag (MPXVgp179) at a variant frequency of 61.1% and a variant P-Value 4.2E-52.

More virulent strains of the virus have been found in isolates from Central Africa (Clade I) compared to Western Africa (Clade II). The differences in amino acids expressed have prompted research into their potential for pathogenicity. The N4R gene has a snp (191565) in which Alanine replaces Threonine. In typical usage, threonine is categorized as a polar hydrophilic (water-loving) amino acid. Protein structure and development rely on its ability to establish hydrogen bonds with a wide range of polar substrates (other molecules). Threonine is a versatile amino acid that can be found either within or on the outside of a protein. It is rather neutral in respect to snp 191565 on N4R gene, though it has a substitution preference with different small or polar amino acids. Alanine is non polar and not hydrophobic.it consists of a normal C-beta carbon and it is present in almost all non-critical protein contexts. Due to the differences in function on the protein structure, it is presumed that the change in amino acid causes snp 191565 on N4R gene to be less virulent. Isoleucine is substituted for Leucine on snp 2723 on J2L gene. Leucine is aliphatic and hydrophobic. It prefers to be entrenched in

protein hydrophobic cores. It also exhibits a fondness for being inside alpha helices more so than in beta strands. Isoleucine prefers to be entrenched in protein hydrophobic cores and it is more laborious for this amino acid to acquire an alpha helical configuration, although it is easy and even preferred for it to stay within beta sheets. Due to the similarity in function of both amino acids on the protein structure, it is presumed that the effect on amino acid causes monkeypox virus to remain the same. Lysine is substituted for glutamate on snp 175877 on BI7R gene Glutamate is charged and polar. It generally prefers to be on the protein's surface, bared to aqueous surroundings. When immersed in the protein, glutamates are usually associated in salt bridges, where they match with a positively charged amino acid to produce stabilizing hydrogen bonds, that can be crucial for stability of protein. Lysine is considered a bit amphipathic (part hydrophilic, part hydrophobic) and are usually associated in salt bridges, where they match with a negatively charged amino acid to produce stabilizing hydrogen bonds that can be crucial for protein stability. Due to the similar function of these amino acids on protein structure, it is presumed that the effect on amin acid remains the same. Asparagine is substituted for Aspartate on snp 176386 on B17R gene. Aspartate is charged and polar and favors being on the protein surface, bared to aqueous surroundings. When immersed within the protein. Aspartates are usually involved in salt bridges, where they match with a positively charged amino acid to produce stabilizing hydrogen bonds, that can be crucial for stability of protein. Asparagine is a polar amino acid which would rather be on the surface of proteins bared to an aqueous environment. Due to the similarity in function of these amino acids on protein structure, it is presumed that the effect on amin acid remains the same. Alanine is nonpolar and not hydrophobic. Nonetheless, it has a normal C-beta carbon, which means it is as hampered as other amino acids with consideration to the conformations that the backbone can acquire. Alanine is substituted by Valine on snp 177993 on B18R gene. Valine is hydrophobic, and prefers to be entrenched in protein hydrophobic cores. Valine contains two non-hydrogen substituents affixed to their C-beta carbon. it is more strenuous for valine to acquire an alpha helical conformation, although it is uncomplicated and even preferred for it to stay within beta sheets. While Alanine and Valine differs in function on protein structure, there is no change to the effect on the amino acid and it remains the same. Glutamate is substituted for glycine on snp 175877 on B17R gene. Glycine is a special amino acid because it contains a hydrogen as its side chain in place of carbon as is the case in other amino acids. This shows that there is considerably more conformational flexibility in glycine. This entails that glycine can inhabit parts of protein structures that are prohibited to all other amino acids. Glutamate is charged and polar. It generally prefers to be on the protein's surface, bared to aqueous surroundings. When

immersed in the protein, glutamates are usually associated in salt bridges, where they match with a positively charged amino acid to produce stabilizing hydrogen bonds, that can be crucial for stability of protein. Due to the similar function of these amino acids on protein structure, it is presumed that the effect on amin acid remains the same.



					Nucleotide	Amino	Acid			Variant	P-			
Position	Reference	Outbreak2022	Type	Ftype	Change	Change		Effect	Protein id	Value		Gene	Locus tag	Product
								Non-synonymous						
2723	А	Т	snp	CDS	A-T	L-I		Variant	NP_536429.1	6.90E-61		J2L	MPXVgp002	J2L
								Non-synonymous						Ankyrin-like
191565	А	G	snp	CDS	A-G	T-A		Variant	NP_536615.1	1.60E-47		N4R	MPXVgp188	protein
								Non-synonymous						
177993	С	Т	snp	CDS	C-T	A-V		Variant	NP_536606.1	4.20E-52		B18R	MPXVgp179	B18R
								Non-synonymous					100	
176386	G	А	snp	CDS	G-A	D-N		Variant	NP_536605.1	4.20E-52		B17R	MPXVgp178	B17R
175877	G	А	snp	CDS	G-A	G-E		Missense Mutation	NP_536605.1	4.20E-52	1	B17R	MPXVgp178	B17R
169783	G	А	snp	CDS	G-A	E-K		Missense Mutation	NP_536599.1	8.60E-76		B11R	MPXVgp172	B11R
54359	С	Т	snp	CDS	C-T	D-N		Missense Mutation	NP_536484.1	9.70E-89		F8L	MPXVgp057	DNA Polymerase
177696	Т	А	snp	CDS	T-A			Synonymous Variant	NP_536605.1	4.20E-52		B17R	MPXVgp178	B17R
175668	А	Т	snp	CDS	A-T			Synonymous Variant	NP_536605.1	4.20E-52		B17R	MPXVgp178	B17R
176526	С	Т	snp	CDS	C-T			Synonymous Variant	NP_536605.1	4.20E-52		B17R	MPXVgp178	B17R
176466	G	А	snp	CDS	G-A			Synonymous Variant	NP_536605.1	2.60E-54		B17R	MPXVgp178	B17R
176130	G	А	snp	CDS	G-A			Silent Mutation	NP_536605.1	4.20E-52		B17R	MPXVgp178	B17R
136139	G	А	snp	CDS	G-A			Silent Mutation	NP_536564.1	3.20E-82		A27L	MPXVgp137	A27L
78-79	CC	TT	snp	CDS				Nonsense Mutation		2.20E-38				
47-49	TTT	AAA	snp	CDS				Nonsense Mutation		3.20E-34				
35-37	TTA	GAG	snp	CDS				Nonsense Mutation		7.60E-33				
22-23	TT	GA	snp	CDS				Nonsense Mutation		1.80E-37				
16-17	AC	TT	snp	CDS				Nonsense Mutation		7.60E-33				
3—4	ТА	AG	snp	CDS				Nonsense Mutation		2.70E-33				
19049	А	Т	snp	CDS				Nonsense Mutation		4.50E-74				
115	С	А	snp	CDS				Nonsense Mutation		8.80E-40				
55	А	Т	snp	CDS				Nonsense Mutation		1.70E-36				
45	Т	А	snp	CDS				Nonsense Mutation		2.00E-32				
40	Т	А	snp	CDS				Nonsense Mutation		5.90E-37				

26	А	С	snp	CDS	Nonsense Mutation	7.60E-33
8	А	Т	snp	CDS	Nonsense Mutation	5.40E-38
178501	G	А	snp	CDS	Nonsense Mutation	4.20E-52
173997	С	Т	snp	CDS	Nonsense Mutation	4.20E-52
102	G	А	snp	CDS	Nonsense Mutation	8.80E-40
44	С	Т	snp	CDS	Nonsense Mutation	2.00E-32
20	А	G	snp	CDS	Nonsense Mutation	7.60E-33
1	G	А	snp	CDS	Nonsense Mutation	5.60E-43
2798	А		snp	CDS	Nonsense Mutation	5.50E-15

Figure 2 shows the table above which is the result of the variants called which include the Position, Reference nucleotides, Variant nucleotides(Outbreak2022), Type, Ftype, Nucleotide change, Amino acid change, Effect, Protein ID, Variant P-value, Gene(J2L,B17R,F8R,A27L,N4R),Locus tag and the products.



3.4 PHYLOGENETIC ANALYSIS

Phylogenetic analysis of all studied sample sequences inclusive of the reference sample was constructed using the National Center for Biotechnology Information (NCBI) [18] and edited using the Interactive Tree of Life (ITOL) [19] to discover the common predecessor of each strain of all 2022 monkeypox virus strains. Comparative analysis of strains was performed on NCBI based on statistical analysis to determine positions of important differentiation between the strains. For each monkeypox virus sequence, we indicated the country, accession number and the date. The result shows that the sequence data generated exhibits various genetic profiles and that random mutations occurred during the evolution process. By applying the clade function, the samples analyzed shows variations going round the tree. The main clades were highlighted with different colours. A clade is a group of organisms that comprises of a similar ancestor and all its lineal descendants. The clades are embedded in each other, as each branch cleave into smaller branches. Of the 372 sequences studied, phylogenesis differentiated the sequences into 12 different clades separated by different colours. The first clade is denoted by the Colour red and has 29 sequences which includes, 10 strains of Portugal, 4 strains of Germany, a strain of Italy, 3 strains of spain, 5 strains of Canada,4 strains of UK, a strain of Switzerland, and a strain of Netherland all separated by their collection dates. The Second clade is represented by the colour purple and has 6 strains of Germany separated by their collection dates. The third clade is denoted by the colour brown and has a total of 27 sequences in the clad of which 21 strains are from Germany,2 strains from Portugal, a strain from Spain, a strain from Switzerland, a strain from USA, and a strain from Canada separated by their collection dates. The fourth clade is represented by a light green colour and has 8 strains of Germany separated by their collection dates. The fifth clade is denoted by a light blue colour and has 10 sequences in total, of which 4 strains are from Germany, 4 strains from Canada, a strain from USA and a strain from Czech Republic all separated by their collection dates. The sixth clade is represented by the colour yellow and has 3 strains from Belgium separated by their collection date. The seventh clade is denoted by the dark green colour and has 3 sequences of which 2 strains are from Germany and a strain from Italy separated by their collection date. The eighth clade is represented by a light purple colour and has 3 sequences of which a strain is from Argentina and 2 strains from Argentina separated by their date of collection. The ninth clade has 2 sequences of which 1 is from Israel and another from USA.it is represented by the colour light blue separated by their collection date. The tenth clade is denoted by the dark green colour and has a total of 4

sequences of which 2 strains are from the UK, a strain from Argentina, and a strain from Australia separated by their date of collection. The eleventh clade is represented by a deep blue colour which has a strain of Slovenia as well as the date collection. The twelfth clade has a total of 264 sequences all separated by their collection dates.6 strains are from Slovenia,111 strains from Germany,33 strains from Portugal,61 strains from Canada,5 strains from Italy, a strain from Argentina, a strain from Australia,7 strains from UK,4 strains from spain,3 strains from Switzerland,4 strains from Belgium,2 strains from Brazil,4 strains from France, a strain from Taiwan, a strain from Mexico, a strain from Sweden,14 strains from USA, a strain from Ireland and 4 strains from Finland all denoted by the light blue colour.[19]

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Figure 6: Phylogenetic tree.

CONCLUSION

Because of the dramatic increase in reported cases, researchers now have access to additional genomes that could provide light on population structure, most notably the potential for multiple introductions of monkeypox virus into the human population. The quantification of variants for monkeypox virus isolates reveals an active strategy for change, but long-term surveillance is required. The increase in the frequency of variants being transferred and introduced from humans to humans may be due to a number of reasons, including inoculation status and the invasion of Human land. Non-human primates and rodents are

prohibited entry due to various countries' strict import regulations. Using data from 376 samples of monkeypox virus in 2022, this study reveals any differences that may have occurred between the sample and reference sequence. For all samples with the identical settings, the variant calling produced around 33 SNPs. We analyzed the effects of polymorphism on protein structure, the types of changes that occurred in amino acids and their implications for the protein, the changes that each codon encodes, the frequency of variations, and the locus tags for each sequence. We went into great detail on the genes affected by variants, the products of those genes, the lengths of the sequences, and the CDS places where changes were found. The collected sequencing data displays a wide range of genetic profiles, suggesting that mutations happened at random during the evolutionary process. Phylogenetic tree construction shows 12 major clades distinguished by colors, each containing creatures with a shared ancestor and their direct genetic descendants. Each clade's branch has split off into subbranches, creating a tree structure. Our research reveals that the evolution of newly emerging strains of monkeypox virus is being influenced by a combination of genomic instability and genetic polymorphism.

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19. Interactive Tree of Life (ITOL)

Supplement materials 1: Summary of monkeypox whole genome sequence isolate investigated. Included are; Accession Number and Demographics

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ISSN 232	20- Aleeession	Sample Name	Data Source	Location	Collection
	Number				Date
	ON800897	ARG003	NCBI	Argentina:	2022-06
				Ciuda	
	ON720962	MPV-ARG002	NCBI	Argentina:	2022-06
				Ciuda	
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ON622720 ON595760 ON918656 OPO19275 OPO19276 OPO19277 ON808413	3ba64538 MPXV-CH-38156923 MPXV-CH-38134631 110-231642 MUW1525179 MUW1527495 MUW1531254 CVR_MPXV1a	NCBI NCBI NCBI NCBI NCBI NCBI	ZHSwitzerlandSwitzerlandSwitzerlandTaiwanAustriaAustriaAustriaUnitedKingdom	2022-05 2022-05 2022-06 2022-07 2022-07 2022-07 2022-05
ON622720 ON595760 ON918656 OPO19275 OPO19276 OPO19277 ON808413	3ba64538 MPXV-CH-38156923 MPXV-CH-38134631 110-231642 MUW1525179 MUW1527495 MUW1531254 CVR_MPXV1a	NCBI NCBI NCBI NCBI NCBI NCBI NCBI	ZHSwitzerlandSwitzerlandSwitzerlandTaiwanAustriaAustriaAustriaUnitedKingdomUnited	2022-05 2022-05 2022-06 2022-07 2022-07 2022-07 2022-05 2022-05
ON622720 ON595760 ON918656 OPO19275 OPO19276 OPO19277 ON808413	3ba64538 MPXV-CH-38156923 MPXV-CH-38134631 110-231642 MUW1525179 MUW1527495 MUW1531254 CVR_MPXV1a	NCBI NCBI NCBI NCBI NCBI NCBI NCBI	ZHSwitzerlandSwitzerlandSwitzerlandTaiwanAustriaAustriaAustriaUnitedKingdomKingdom	2022-05 2022-05 2022-06 2022-07 2022-07 2022-07 2022-05 2022-05
ON622720 ON595760 ON918656 OPO19275 OPO19276 OPO19277 ON808413 ON808414	3ba64538 MPXV-CH-38156923 MPXV-CH-38134631 110-231642 MUW1525179 MUW1527495 MUW1531254 CVR_MPXV1a CVR_MPXV1b CVR_MPXV1c	NCBI NCBI NCBI NCBI NCBI NCBI NCBI	ZHSwitzerlandSwitzerlandSwitzerlandTaiwanAustriaAustriaAustriaUnitedKingdomKingdomUnitedUnited	2022-05 2022-05 2022-06 2022-07 2022-07 2022-07 2022-05 2022-05
ON622720 ON595760 ON918656 OPO19275 OPO19276 OPO19277 ON808413 ON808414	3ba64538 MPXV-CH-38156923 MPXV-CH-38134631 110-231642 MUW1525179 MUW1527495 MUW1531254 CVR_MPXV1a CVR_MPXV1b	NCBI NCBI NCBI NCBI NCBI NCBI NCBI	ZHSwitzerlandSwitzerlandSwitzerlandTaiwanAustriaAustriaAustriaAustriaKingdomUnitedKingdomUnitedKingdomKingdomKingdom	2022-05 2022-06 2022-07 2022-07 2022-07 2022-05 2022-05 2022-05
ON622720 ON595760 ON918656 OPO19275 OPO19276 OPO19277 ON808413 ON808415 ON808416	3ba64538 MPXV-CH-38156923 MPXV-CH-38134631 110-231642 MUW1525179 MUW1527495 MUW1531254 CVR_MPXV1a CVR_MPXV1b CVR_MPXV1c	NCBI NCBI NCBI NCBI NCBI NCBI NCBI	ZHSwitzerlandSwitzerlandSwitzerlandTaiwanAustriaAustriaAustriaAustriaKingdomUnitedKingdomUnitedKingdomUnited	2022-05 2022-06 2022-07 2022-07 2022-07 2022-07 2022-05 2022-05 2022-05

ON808417	CVR_MPXV1e	NCBI	United	2022-05
			Kingdom	
ON645312	MPXV_GSTT_Patient1	NCBI	United	2022-05
			Kingdom	
ON619835	MPXV_UK_2022_1	NCBI	United	2022-05
			Kingdom	
ON619836	MPXV_UK_2022_2	NCBI	United	2022-05
			Kingdom	
ON619837	MPXV_UK_2022_3	NCBI	United	2022-05
			Kingdom	
ON619838	MPXV_UK_2022_4	NCBI	United	2022-05
			Kingdom	
ON959131	IL001	NCBI	USA:IL	2022-05
ON959132	IL003	NCBI	USA:IL	2022-06
ON959133	NY002	NCBI	USA: NY	2022-05
ON959134	NY003	NCBI	USA: NY	2022-05
0N959135	NY004	NCBI	USA: NY	2022-05
ON959136	NY005	NCBI	USA: NY	2022-06
ON954773	CA002	NCBI	USA:CA	2022-05
ON674051	MPXV-USA-2022-	NCBI	USA: FL	2022-05
	FL001			
ON675438	MPXV-USA-2022-	NCBI	USA:VA	2022-05
	VA001			
ON676703	MPXV-USA-2022-	NCBI	USA:CA	2022-05
	CA001			
ON676704	MPXV-USA-2022-	NCBI	USA: FL	2022-05
	FL002			
ON676705	MPXV-USA-2022-	NCBI	USA: UT	2022-05
	UT001			
ON676706	MPXV-USA-2022-	NCBI	USA: UT	2022-05
	UT002			

OPO18607	RKI185	NCBI	Germany	2022-07
OPO18606	RKI184	NCBI	Germany	2022-07
ON627808	UT-UPHL-82200022	NCBI	USA	2022-05
ON563414	MPXV-USA-2022-	NCBI	USA:MA	2022-05
	MA001			

Supplement Materials 2;

SNP Analysis with no Effect on Protein

Based on the 33 results gotten from variant analysis between the 387 samples mapped to Zaire-96-I-16 as reference in this study,6 of the variants encodes changes in amino acid but have no effects on amino acid. This is a Silent mutation, which occurs when a change in the DNA sequence of a gene does not affect the subsequent amino acid sequence of the protein it codes for. This is shown in Fig 4. 20 of the variants have no effects on amino acid and therefore do not encode changes. This is a Nonsense mutation, which occurs in DNA when a sequence change gives rise to a stop codon rather than a codon specifying an amino acid. This is shown in Fig 5.

SNP136139, with variant G, as reference nucleotide and A as the variant nucleotide (Guanine-Adenine) encodes a codon change (AGC-AGT) resulting to no effect in the amino acid with protein id (NP_536564.1), a length of 1 and locus tag (MPXVgp137) at a variant frequency of 82.1% and a variant P-Value 3.2E-82. SNP175668, with variant A as reference nucleotide and T as variant nucleotide (Adenine-Thymine) encodes a codon change (ACA-ACT) resulting to no effect in the amino acid with protein id (NP_536605.1), a length of 1 and locus tag (MPXVgp178) at a variant frequency of 61.1% and a variant P-Value (approximate) 4.2E-52. SNP176130, with variant G, as reference nucleotide and A as variant nucleotide (Guanine-Adenine) encodes a codon change (GTG-GTA) resulting to no effect in the amino acid with protein id (NP_536605.1), a length of 61.1% and a variant P-Value (approximate) 4.2E-52. SNP176130, with variant G, as reference nucleotide and A as variant frequency of 61.1% and a variant P-Value (approximate) 4.2E-52. SNP176466, with variant G, as reference nucleotide and A as variant frequency of 61.1% and a variant nucleotide (Guanine-Adenine) encodes a codon change (GTG-GTA) resulting to no effect in the amino acid with protein id (NP_536605.1), a length of 1 and locus tag (MPXVgp178) at a variant frequency of 61.1% and a variant P-Value (approximate) 4.2E-52. SNP176466, with variant G, as reference nucleotide and A as variant nucleotide (Guanine-Adenine) encodes a codon change (GTG-GTA) resulting to no effect in the amino acid with protein id (NP_536605.1), a length of 1 and locus tag (MPXVgp178) at a variant frequency of 61.1% and a variant P-Value (approximate) 4.2E-52. SNP176466, with variant G, as reference nucleotide and A as variant nucleotide (Guanine-Adenine) encodes a codon change (GTG-GTA) resulting to no effect in the amino acid

with protein id (NP 536605.1), a length of 1 and locus tag (MPXVgp178) at a variant frequency of 61.1% and a variant P-Value (approximate) 2.6E-54. SNP176526, with variant C, as reference nucleotide and T as variant nucleotide (Cytosine-Thymine) encodes a codon change (CAC-CAT) resulting to no effect on the amino acid with protein id (NP_536605.1), a length of 1 and locus tag (MPXVgp178) at a variant frequency of 61.1% and a variant P-Value (approximate) 4.2E-52. SNP177696, with variant T, as reference nucleotide and A as variant nucleotide (Thymine-Adenine) encodes a codon change (GTT-GTA) resulting to no effect in the amino acid with protein id (NP_536605.1), a length of 1 and locus tag (MPXVgp178) at a variant frequency of 61.1% and a variant P-Value (approximate) 4.2E-52.SNP 1 with G, as reference nucleotide and A as variant nucleotide (Guanine-Adenine) encodes no codon change resulting in no effect to amino acid with no protein id, a length of 1 and no locus tag at a variant frequency of 80.0% and a variant P-Value 5.6E-43. SNP 3-4 with TA, as reference nucleotide and AG as variant nucleotide (Thymine Adenine- Adenine Guanine) encodes no codon change resulting in no effect to amino acid with no protein id, a length of 2 and no locus tag at a variant frequency of 66.7% and a variant P-Value 2.7E-33.SNP 8 with A as reference nucleotide and T as the variant nucleotide (Adenine-Thymine)) encodes no codon change resulting in no effect to amino acid with no protein id, a length of 1 and no locus tag at a variant frequency of 73.3% and a variant P-Value 5.4E-38. SNP 16-17 with AC as reference nucleotide and TT as variant nucleotide (Adenine Cytosine-Thymine Thymine) encodes no codon change resulting in no effect to amino acid, no protein id, length of 2 and no locus tag at a variant frequency of 64.5% and a variant P-Value 7.6E-33.SNP 20 with A as reference nucleotide and G, as variant nucleotide (Adenine-Guanine) nucleotide encodes no codon change resulting in no effect to amino acid, no protein id, a length of 1 and no locus tag at a variant frequency of 64.5% and a variant P-Value 7.6E-33. SNP 22-23 with TT as reference nucleotide and GA as variant nucleotide (Thymine Thymine-Guanine Adenine) encodes no codon change resulting in no effect to amino acid, no protein id, length of 2 and no locus tag at a variant frequency of 71.0% and a variant P-Value 1.8E-37. SNP 26 with A as reference nucleotide and C as variant nucleotide (Adenine-Cytosine) encodes no codon change resulting in no effect to amino acid, no protein id, length of 1 and no locus tag at a variant frequency of 64.5% and a variant P-Value 7.6E-33. SNP 35-37 with TTA as reference nucleotide and GAG as variant nucleotide (Thymine Thymine Adenine- Guanine Adenine Guanine) encodes no codon change resulting in no effect to amino acid, no protein id, length of 3 and no locus tag at a variant frequency of 64.5% and a variant P-Value 7.6E-33. SNP 40 with T as reference nucleotide and A as variant nucleotide (Thymine-Adenine) encodes no codon change resulting in no effect to amino acid and therefore no protein id, a length of 1 and no locus tag at a variant frequency of 68.8% and a variant P-Value 5.9E-37. SNP 44 with C as reference nucleotide and T as variant nucleotide (Cytosine-Thymine) encodes no codon change resulting in no effect to amino acid, no protein id, with a length of 1 and no locus tag at a variant frequency of 62.5% and a variant P-Value 2.0E-32. SNP 45 with T as reference nucleotide and A as variant nucleotide (Thymine-Adenine) encodes no codon change resulting in no effect to amino acid, no protein id, a length of 1 and no locus tag at a variant frequency of 62.5% and a variant P-Value (approximate) 2.0E-32. SNP 47-49 with TTT as reference nucleotide and AAA as variant nucleotide (Thymine Thymine Thymine-Adenine Adenine) encodes no codon change resulting in no effect to amino acid, no protein id, length of 3 and no locus tag at a variant frequency of 60.6% and a variant P-Value 3.2E-34. SNP 55 with A as reference nucleotide and T as variant nucleotide (Adenine-Thymine) encodes no codon change resulting in no effect to amino acid, no protein id, a length of 1 and no locus tag at a variant frequency of 66.7% and a variant P-Value 1.7E-36. SNP 78-79 with CC as reference nucleotide and TT as variant nucleotide (Cytosine Cytosine-Thymine Thymine) encodes no codon change resulting in no effect to amino acid, no protein id, length of 2 and no locus tag at a variant frequency of 61.5% and a variant P-Value 2.2E-38. SNP 102 with G as reference nucleotide and A as variant nucleotide (Guanine-Adenine) encodes no codon change resulting in no effect to amino acid, no protein id, a length of 1 and no locus tag at a variant frequency of 61.0% and a variant P-Value 8.8E-40. SNP 115 with C as a reference nucleotide and A as variant nucleotide (Cytosine-Adenine) encodes no codon change resulting in no effect to amino acid, no protein id, a length of 1 and no locus tag at a variant frequency of 61.0% and a variant P-Value 8.8E-40. SNP 2798 has a deleted reference nucleotide and A as a variant nucleotide (-Adenine) encodes no codon change resulting in no effect to amino acid, no protein id, a length of 1 and no locus tag at a variant frequency of 64.7% and a variant P-Value 5.5E-15. SNP 19049 with A as a reference nucleotide and T as a variant nucleotide (Adenine-Thymine) encodes no codon change resulting in no effect to amino acid, no protein id, a length of 1 and no locus tag at a variant frequency of 72.1% and a variant P-Value 4.5E-74. SNP 173997 with C as a reference nucleotide and T as a variant nucleotide (Cytosine-Thymine) encodes no codon change resulting in no effect to amino acid, no protein id, a length of 1 and no locus tag at a variant frequency of 61.1% and a variant P-Value 4.2E-52. SNP 178501 with G, as a reference

nucleotide and A as a variant nucleotide (Guanine-Adenine) encodes no codon change resulting in no effect to amino acid, no protein id, a length of 1 and no locus tag at a variant frequency of 61.1% and a variant P-Value 4.2E-52.

Figure 4. Silent Mutation



SNP 136139 shows the codon change on the coding region of the A27L gene.



Figure 5 Nonsense Mutation

SNP 3-5 shows no effect in amino acid and shows a stop codon.