



Revitalization of plant growth promoting rhizobacteria for sustainable development in agriculture.

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ABSTRACT

PGPR are those bacteria occurring as natural colonizers of soil which had gained worldwide importance in the field of agricultural enhancement. They had been observed in essential requirements to increase the productivity of soil by regaining fertility. Advancement of life in all systems has been nowadays not only reliant on farming but diets safety play a chief role in satisfying the growing population basic needs. The viability of soil environments serve as a source of a non-stop manifestation of soil bacteria. There are even other aspects of soil

which play conservatory role in soil fertility; including of plants in synergistic co-evolution, soil microbes and bio-mineralization. The growth of population globally has put pressure on farming thus demanding chemical fertilizer's high yield. Meanwhile with the application of peats and insect killer in the farming area have ruined the soil worth and richness, resulting in contraction of farming acreage having productive soil, therefore the consideration of scientists and researchers has moved towards harmless and fruitful sources of farming carry out. (PGPR) Plant growth promoting rhizobacteria has been effective by co-evolution between plants and microbes. Bacteriological revival had been attained by plant growth promoters activated by direct and indirect tactics such as disease resistance, rhizoremediation, bio-fertilization, invigorating root growth, etc. The sort of unpredictability existed in the working of PGPR because of several ecological influences that effect their progression and propagation in plants. Due to existent limitations playing its role in agronomy PGPR applications were not improving. These restrictions could be overcome by usage of present methodologies and practices such as Micro-encapsulation and Nano-encapsulation. The introduction of new coming and modern research techniques are supporting their applications with the help of fields such as chemical engineering, biotechnology, agro biotechnology, nanotechnology, and material science by bringing together different environmental and practical living approaches to arrange new designs and prospects of huge prospective.

Keywords: Agriculture Biofertilization Nano-encapsulation PGPR Revitalization Sustainable development

INTRODUCTION

Farming remained the biggest economic cause standing from the time of beginning of advancement. Around 7.41 billion people dwelling around globe, occupy 6.38 billion hectares of world area, out of which 1.3 billion people are almost reliant on cultivation. With the increasing urge for food, soil viability is primarily significant for conservatory aspect of cultivation (Paustian *et al.*, 2016; Tschardt *et al.*, 2012). Crop growing Association of the United Nations termed as (FAO) provided Food Balance Sheet 2004 which lead to conclusion that 99.7% of foodstuff for the population globally originated from terrestrial location only. A food fact suggested that 79 million people are newly added to the global population annually, there is no doubt; a constant rise in food price which resulted in a concurrent unavailability in resources (Alexandratos and Bruinsma, 2003). In India, 60.6% of

land is used for cultivation which serve as a driving force from half of its population to raise varieties of pulses vegetables and cereals. Three chief factors like Farming outputs, water class, and climate variability are most prominently effected by the exchange of atmosphere, the aquatic ecosystem, energy, the soil environment and carbon resources within soil organic materials (Lehmann and Kleber, 2015). Various constituents of soil which marks the wealthy nature vital for soil richness include abiotic and biotic factors, organic carbon content, nutrients, moisture, nitrogen, potassium and phosphorous. Yet, the major issue which brings negativity is unselective use of composts, mainly phosphorus and nitrogen, which lead to extensive soil effluence through exchangeable bases and reducing pH. Consequently, these are the phenomenon's responsible for nutrients unreachable to crops causing productivity loss which could increase if not controlled (Gupta *et al.*, 2015). According to FAO, 38.47% of the global acreage space is enclosed by cultivated area, of which 28.43% accounts for arable land necessary for cultivation. In accordance with record of world food and agriculture authority; lone 3.13% is always required for cultivation. Furthermore, the conditions are worsened when; 20–25% of land globally is continually ruined every year and extra 5–10 Mha, is unavoidably ruined each year which is nowhere getting controlled (Abhilash *et al.*, 2016). Since developing farming area need proper management, because extraordinary demand puts grave burden upon terrestrial environment for extraordinary yield. Therefore, an extra advanced and scientific agricultural practice is needed for satisfying the rising loads that are useful conserving the soil productiveness. Nearly some of the modern techniques participating in effective farming termed as practical running carry out (Ubertino *et al.*, 2016), agricultural intensification (Shrestha, 2016), bio fertilizers usage (Suhag, 2016; Kamkar, 2016), use of microbes or genetically engineered microbes to support growth of plant (Perez *et al.*, 2016; Kumar, 2016), genetically engineered crops to form nitrogen-fixing symbioses, and fixing nitrogen in the absence of microbial symbionts (Mus *et al.*, 2016; Passari *et al.*, 2016). Moreover, there are several new scientific and socioeconomic practices which participate in viable agricultural improvement include salt tolerance, drought tolerance, disease resistance, better nutritional value, and heavy metal stress tolerance.

Nowadays as the modern world is advancing, it brings new hopes for overcoming these negativities. The latest technique implies the usage of soil microbes, for example bacteria, fungi, and algae, which are quite promising mode to satisfy these ideal objectives (Vejan *et al.*, 2016). Microorganisms and leguminous plants having holobiont relationships by synergistic co-evolution and bio-mineralization have remarkable capacity for refining fertility and

quality of soil (Paredes and Lebeis, 2016; Rosenberg and Rosenberg, 2016; Agleret *al.*, 2016). soil microbes having Co-evolution with plants is important technology to counterlife-threatening abiotic environments, leading to enhanced environmental sustainability, economic viability, and soil fecundity (Khan *et al.*, 2016; Compant *et al.*, 2016). plant growth promoting rhizobacteria (PGPR) has no doubt truly illuminated the relationship between microbes and plants, which is an indicator of synergistic and antagonistic relations causing upgrading of plant growth (Rout and Callaway, 2012; Bhardwaj *et al.*, 2014). PGPR soil physiognomies are hugely effected by PGPR which proves an important role in transforming poor quality, unfertile area into fertile acreage. Regeneration of growth of plant and quality of soil through PGPR had been a field vigorously misused for improved agricultural yield in many areas globally (Gabriela *et al.*, 2015). This task is commonly accomplished through indirect or direct grounds. This is directly achieved by direct provision of plants compounds that enhance growth of plant. This availability is reachable through methods for example plant stress control, bio-fertilization, and rhizo-remediation (Goswami *et al.*, 2016). Sucking nutrients and water from the soil is generally the most fundamental ecological element limiting development of terrestrial plant species. PGPR like bio-fertilization enhances growth of plant through increase in the availability or suction of nutrients from a restricted soil nutrient land. Counteracting stress of plant is additional vital PGPR affect and is applicable equally to abiotic and biotic stress. It has been proved that biotic stress is a living danger (disease, insects), however abiotic stress which could be physical (temperature, light etc.) or chemical stress that the surroundings put upon plants (Gabriela *et al.*, 2015) are equally harmful for the adequate growth of plant. PGPR also indirectly participate in improvement of plant growth by subsidizing or stopping the toxic effects of one or more phyto-pathogenic organisms. In the present instance, growth of plant is enhanced through mechanisms like competitive exclusion, antibiosis and induction of systemic resistance (ISR) (Tripathi *et al.*, 2012).

Since results need an improvement in the field of plant growth by PGPR, due to lack of information amongst its present uses and the possible PGPR applications for ecological growth. The field of applications of PGPR is moreover extremely restricted because of unpredictability and irregularity in results perceived in research laboratory, field trails and green house. Although these problems can be solved by means of present nanobiotechnological methodologies and use of procedures for example micro-encapsulation and nano-encapsulation. These outcomes signifies only few methodologies which can be

revised for PGPR implementation as an instrument to fight plant infections and boost agricultural yield.

PLANT GROWTH ENHANCING RHIZOBACTERIA

Plants in their phase of progression and growth involve symbiotic relationship with soil microorganisms (fungus and bacteria). The free-living microbes of soil living in rhizosphere of various species of plant; have considerably varied useful results on the host plant (Raza *et al.*, 2016a,b) by implementation of dissimilar mechanisms considered as (PGPR) Plant Growth Promoting Rhizobacteria for instance nodulation and nitrogen fixation. They are helpful in guarding plant's vigour in an environmentally friendly way (Akhtar *et al.*, 2012). PGPR and their plant interactions are implied in scientific practices but nowadays in this modern era they are constantly misused for the support of commercial and agricultural purposes (Gonzalez *et al.*, 2015). These interactions include applications examined on cucumber, radicchio, wheat, barley, lentil, oat, peas, tomato, maize, canola, potato, and soy (Gray and Smith, 2005). PGPR is involved in countless viable approaches in the soil environment to make it vigorous for the profitable purpose and they are no doubt justified for crop yield (Gupta *et al.*, 2015). Their compatibility to inhabit roots system of plants and improve growth of plant is accomplished through varied techniques, including degradation of environmental pollutants, hydrogen cyanate (Liu *et al.*, 2016); siderophores (Jahanian *et al.*, 2012), 1-amino-cyclopropane-1-carboxylate (ACC) deaminase, production of indole-3-acetic acid (IAA), antibiotics or lytic enzymes (Xie *et al.*, 2016), production of hormones, phosphate solubilization (Ahemad and Khan, 2012) nitrogen fixation (Glick, 2012). Moreover, Since PGPR is famous promoter for qualitative plant growth, therefore they are necessary and thus applicable on insects, biological control of phytopathogens, salinity tolerance and heavy metal detoxifying activities (Egamberdieva and Lugtenberg, 2014).

2.1. Rhizosphere

Rhizosphere are referred as microorganism store room colonizing soil area neighbouring to roots of plants and thus helpful in maintaining soil biotic and chemical characteristics. Bacteria resident in rhizosphere can be symbiotic or non-symbiotic, consolidated through determining their useful and harmful modes of action (Kundan *et al.*, 2015). The function of root system is anchorage and sucking of nutrients and water from soil, which is a considered fact implied in chemical factory wherever phenolic compounds are generated and instantaneously released to mediate several underground interactions. The plant root's compounds are constantly released

servicing chemical attractants for enormous but diverged microscopic groups. These compounds include composition influenced by the physical position and plant's species microbes (Kang *et al.*, 2010). Three dissimilar constituents' form the rhizosphere are: the root, the rhizoplane, and the rhizosphere itself. Among them, the rhizosphere is present in area of soil controlled by plant's root with the help of substrate secretion and generalized infectious action. The function of rhizoplane present on the root surface is to powerfully fix with particles of soil, where root has been occupied with microbes (Barea *et al.*, 2005). The bacterial concentration dwelling in rhizosphere is almost 10–1000 times higher as comparatively to soil greater part, but lesser as compared to research laboratory medium. In order to conserve their useful outcomes within the root environs, there is a demand of compatibility with more rhizosphere microorganisms for the purpose of food released specifically by the root. The interactions between the rhizosphere and plant are vital to suck nutrients and water from soil and these interactions are fruitful to both the soil-borne microbes and plants.

2.2. Different types of PGPR

The classification of PGPR is recognized and divided into two chief types namely intracellular plant growth promoting rhizobacteria (iPGPR) and extracellular plant growth promoting rhizobacteria (ePGPR) (Viveros *et al.*, 2010). ePGPR live in the gaps amongst cells of the root cortex or rhizosphere (on the rhizoplane), However iPGPR principally dwell within particular nodes of root cells. The genera of bacteria referred as ePGPR are Burkholderia, Pseudomonas, Micrococcus, Arthrobacter, Azotobacter, Erwinia, Agrobacterium, Serratia, Chromobacterium, Flavobacterium, Caulobacter, Azospirillum, and Bacillus. The iPGPR endophytic microorganisms are Frankia species, Rhizobium, Mesorhizobium and Bradyrhizobium, that has the capacity to fix nitrogen in air particularly in higher plants (Bhattacharyya and Jha, 2012).

PGPR ROLE IN GROWTH AS PLANT PROMOTER

PGPR improve growth of plant through particular qualities (Gupta *et al.*, 2015). These qualities of PGPR include increase in growth of plant with the help of indirect and direct procedures, like phytopathogens resistance and improvement in physiology of plants, by varied actions and approaches (Zakry *et al.*, 2012). Actions are widespread approaches which include enzymes for disease prevention, producing volatile organic compounds (VOCs), neutralizing abiotic and biotic stress and nutrient fixation. Though, the type of action by

various PGPR types differs depending on plant host type (Garcia *et al.*, 2015). They are also effected by a various biotic factors (climatic conditions, further microbial community members, plant defence mechanisms, plant developmental stages and plant genotypes) and abiotic factors (management and composition of soil) (Vacheron *et al.*, 2013). Biotic and abiotic approaches are two important and vital foundations for PGPR applications in plant growth and nevertheless both are fundamental and necessary part of story

DIRECT TOOLS

PGPR is responsible for direct facilitation of the development and growth of plants with application of procedures like phytohormones production, solubilisation of mineral nutrients, mineralization of organic compounds, rise in nutrient obtainability by nitrogen fixation or uptake of nutrients (Bhardwaj *et al.*, 2014). These mechanisms are very fruitful and thus responsible for direct plant growth which is entirely dependent on the plant species and type of microbial strain. Enhancement of mineral suction from the soil through root surface is directly dependent on singular ion flux and this is accomplished by the PGPR existence.

3.1.1. FIXING OF NUTRIENT

PGPR serve as direct growth promoters in plants, since they are linked with capacity which increase the concentration and availability of nutrients by locking or fixing the plant growth supply and yield (Kumar, 2016). Plants suck soil nitrogen in the form of ammonium (NH_4^+) and nitrate (NO_3^-), and thus referred as essential nutrients for development. Nitrate is known as the predominant form of nitrogen supply in aerobic soils through plants where nitrification prominently take place (Xu *et al.*, 2012). Some PGPR have the capacity of phosphate solubility, causing a better amount of phosphate ions accessible from the soil, which can be definitely absorbed by plants (Paredes and Lebeis, 2016). *Kocuria Turkanensis* 2M4 isolated from the soil rhizosphere serve as an IAA producer for various plant species. It is also act as siderophore producer, and phosphate solubilizer (Goswami *et al.*, 2014). Additionally microbes such as *Bacillus subtilis* UPMB10, *Acinetobacter* sp. S3r2, *Bacillus pumilus* S1r1, *Klebsiella pneumoniae* Fr1, and *Klebsiella* sp. Br1 have the area potential for fixing atmospheric N_2 , and delay N_2 remobilization.

FIXING OF NITROGEN

Biotic way of nitrogen fixation is an astonishing process that explains for almost two-thirds of the nitrogen fixed universally. This organic method is executed either by symbiotic or non-

symbiotic interactions amongst plants and microbes (Shridhar, 2012). Symbiotic PGPR, which are most commonly caused the fixation of atmospheric N^2 in soil, contain strains of *K. pneumoniae*, *Pantoea agglomerans*, *Beijerinckia* sp., *Azoarcus* sp., and *Rhizobium* sp., (Ahemad and Kibret, 2014). Inoculation of several rhizobacterial species into soil enhances its quality and increases nodule formation. A particular gene called *nif* is involved in undertaking N^2 fixation, while additional structural gene is involved in switching on the iron protein, biosynthesizing the iron molybdenum cofactor, donating electrons and numerous other regulatory genes compulsory for the enzyme's activity and synthesis (Reed *et al.*, 2011). Biological N^2 -fixing through PGPR is an authenticated process perceived by inoculation on agricultural fields and crops and thus fruitful in reviving activity by building growth enhancement. It is an extraordinary source that retains the level of nitrogen in crop soil and play role in disease management (Damam *et al.*, 2016).

3.1.3. SOLUBILIZATION OF PHOSPHATE

Phosphorus is the second most necessary nutrient compulsory for plants in suitable amounts for ideal growth. It serve as key part in nearly all main metabolic processes, consist of photosynthesis, macromolecular biosynthesis, energy transfer, respiration and signal transduction (Anand *et al.*, 2016). However, 95–99% of phosphorus existent in precipitated, immobilized, or insoluble forms consequently, making its uptake difficult for plants. The uptake of phosphate in plants occur lone as dibasic (HPO_4^{2-}) and monobasic ($H_2PO_4^-$) ions. Phosphate-solubilisation is the only process implied by bacteria still involved in mineralization and Solubilisation of phosphorus and is attained chiefly through PGPR. The organic acids of low molecular weight are synthesized by countless bacteria's of soil, responsible for inorganic phosphorus solubility which are no doubt miraculous approach effective in agriculture side (Sharma *et al.*, 2013). Phosphate solubility in PGPR are provided by the bacterial genera of *Serratia*, *Rhodococcus*, *Enterobacter*, *Flavobacterium*, *Mesorhizobium*, *Rhizobium*, *Pseudomonas*, *Arthrobacter*, *Erwinia*, *Microbacterium*, *Bacillus*, *Burkholderia*, and *Beijerinckia*, that has appealed to agriculturists thus confirming this fact that inoculation of soil increase plant growth and production (Oteino *et al.*, 2015). Amongst them, *Mesorhizobium mediterraneum* and *Mesorhizobium ciceri* isolated from nodules of chickpea, are worthy phosphate solubilizers (Parmar and Sindhu, 2013). No doubt nowadays these microorganisms are fruitful and definitely needed for phosphorus solubility causing an improvement in soil fertility. But we cannot deny this fact that still in spite of extensive research; results are inadequate regarding its application as a bio-fertilizers.

3.1.4. SOLUBILIZATION OF POTASSIUM

The third necessary macronutrient for the growth of plants is Potassium. Since greater than 90% of potassium is available in the form of silicate minerals and insoluble rock but still concentration of soluble potassium is nevertheless very small in soil (Parmar and Sindhu, 2013). Aforemost limitation implying in grooming agricultural yield is potassium deficiency. In the absence of sufficient potassium, proven poor growth of roots, lesser yield, ceased growth rate, and low seed yield has resulted. There is an inevitable demand required to search a substantial endemic potassium source for keeping potassium wealth and for much better absorbing capacity for adequate crop yield (Kumar and Dubey, 2012). The PGPR capacity to solubilize potassium rock through synthesizing and releasing organic acids has been broadly explored. Potassium solubilizing PGPR, for example Burkholderia., Pseudomonas sp., Bacillus mucilaginosus., Paenibacillus sp., Ferrooxidans sp., Acidithiobacillus sp and Bacillus edaphicus sp., are employed in providing amazing results to secrete potassium in reachable form through potassium-bearing minerals existing in soil (Liu *et al.*, 2012). Consequently, with the application of potassium-solubilizing PGPR serving as bio fertilizer for the fruitful farming can decrease the surge of agrochemicals needs and better upkeep of agricultural yield proven in environmental friendly way (Setiawati and Mutmainnah, 2016).

3.1.5. PHYTOHORMONE RELEASE

Phytohormones or plant growth regulators are referred as biological substances, which are found at relatively low concentrations (<1 mM) modify, promote or inhibit plant's development and growth (Damam *et al.*, 2016). Luckily, yield of these phytohormones can even be induced through certain microorganisms, for example PGPR, in plants. Generally groups of phytohormones important and released from PGPR source consist of auxins, brassinosteroids, gibberellins, ethylene, abscisic acid and cytokinins. With the arrival of phytohormones in the agricultural field, a considerable rise in water and food absorbance in root cell is observed which can even multiply by overproducing root hairs and lateral roots (Sureshbabu *et al.*, 2016). Plant growth regulators are recently known which are referred as exogenous plant hormones, since they are applicable exogenously as synthetic analogues or extracted hormones onto plant tissues or whole plants. Phytohormones are classified by their dependence on area of **action**.

(A) INVIGORATION OF ROOT

Invigoration of root consist of various hormone-mediated pathways which coincide with pathways that are identified as exterior (Jung *et al.*, 2013). These hormones involve production in a way that sometimes involvedefinite microorganisms, such as *Rhizobium leguminosarum*, *Azotobacter chroococcum*, *Klebsiella oxytoca*, *Pseudomonas putida*, *Stenotrophomonas maltophilia*, *Pseudomonas aeruginosa*, *Mesorhizobium ciceriand*, *Enterobacter asburiae* and *Paenibacillus polymyxa*, which are considered as PGPR. Hormones for exampleethylene, kinetin, gibberellins, and auxins are specifically formed by these microorganisms and serve a vital role in invigoration of root(Ahemad and Kibret, 2014).

(B) INVIGORATION OF SHOOT

The hormones such as auxins, gibberellins and Cytokinins, are important growth hormones that control all phases of growth in higher plants. Skoog and Miller, 1957 analysed that if we increase hormone availability; a positive relationship is built between cytokinins concentration and shoot development instead of root development. Few foremost cytokinins includedihydrozeatin [6(4-hydroxy-3-methyl-butylamino) purine], cis-zeatin [6-(4hydroxy-3-methyl-cis-2-butenylamino) purine], trans-zeatin [6(4-hydroxy-3-methyl-trans-2-butenylamino) purine], and i6Ade [6-(3-methyl-2-butenylamino) purine] (Murai, 2014). The smooth making and assemblage of these hormones of plants through microorganisms could serve as an energetic attempt for positive changein agricultural yield and help in getting advancedand preferred traits. Microorganisms that serve as a resource in hormones assemblage, are even involved in application of shoot invigoration that is mostly observed in PGPR, like *Azotobactersp*, *Pseudomonas sp.*, *Paenibacillus polymyxa*, *Rhizobium leguminosarum*, *Bacillus subtilis*, *Pantoeaagglomerans*, *Pseudomonas fluorescens*, and *Rhodospirillum rubrum* (Prathap and Ranjitha, 2015).

3.1.6. SIDEROPHORE RELEASE

Siderophores are referred as small biological molecules obtained by microbes in iron-limiting conditions which support and improve iron absorbing potential. The results of Research on siderophores has concluded that during recent decadethe iron metal ion absorbance capacity has developed (Saha *et al.*, 2016). The plant microbe such as *Pseudomonas sp.*, which serve as PGPR, consumes the siderophores takenby rhizosphere microorganisms for satisfying their demand of iron. Particularly, the microbe such as *Pseudomonas putida* consumes heterologous siderophores justified from other microbes in

order to improve the iron availability level within the ecosystem (Rathore, 2015). An effective siderophore, for example the complex of ferric-siderophore, serves a vital part in plant's iron absorbance in the presence of metals, like cadmium and nickel (Beneduzi et al., 2012). Since PGPR is known for production of siderophores, thus serving as a foremost part in satisfying iron demand. The research conducted on siderophores potential about rise in iron absorbance is till now very restricted, therefore further research is demanded in this field.

3.1.7. EXOPOLYSACCHARIDE RELEASE

Exopolysaccharides (EPSs) are included in a group regarding biodegradable polymers having high molecular weight involving derivatives and monosaccharide residues from various plants, algae and bacteria (Sanlibaba and Cakmak, 2016). EPSs play a pivotal role in chief part in survival of host in stress conditions like (water logging, saline soil, or dry weather) or pathogenesis and therefore are suggested for good agricultural yield through obligate interaction amongst rhizobacteria and plant roots, soil particle's aggregation and maintenance of water potential (Pawar *et al.*, 2016). EPS results confirmed the production of PGPR, for instance from *Rhizobium* sp., *Xanthomonas* sp., *Agrobacterium* sp., *Enterobacter cloacae*, *Bacillus drentensis*, *Rhizobium leguminosarum* and *Azotobacter vinelandii*, are very important to capacitate a rise in fertility of soil by supporting agricultural yield (Mahmood *et al.*, 2016).

3.1.8. BIOLOGICAL-FIXATION OF ATMOSPHERIC NITROGEN

Atmospheric nitrogen fixation is referred as non-mutualistic or mutualistic association between plants providing a fixed carbon and suitable habitat by microorganisms through fixed nitrogen exchange (Kuan *et al.*, 2016). The mentioned association amongst plants and PGPR is observed in the genera *Pseudomonas*, *Bacillus*, *Azospirillum*, *Burkholderia*, and *Klebsiella* which has been extensively studied (Islam *et al.*, 2016). But, still these practices are limited mostly to legumes in agrarian sciences because of lack of substantial curiosity existing in the field of discovery of either symbiotic or non-symbiotic relationship between non-legumes.

3.1.9. RHIZOREMEDIATION

Contamination of water and soil has been a critical issue emerging globally these days. The major type of problems arising nowadays is a lethal source of pollution in ecosystem. Pollution can be lessened through bioremediation, referred as a method or system in which living entities or their goods are used artificially or naturally to immobilize, destroy or remediate

pollutants in the ecosystem (Uqab *et al.*, 2016). Since bioremediation is known to be an economical and time taking resource harboured for tackling pollution of water and soil. The several bioremediation methods are existent, including bioventing, bio-slurry, bio-pile, phytoremediation and landfarming. All these techniques has been applicable in polluted areas for to cut down waste products. In spite of several advancements, yet; while applying these procedures we need to execute a relationship which is applicable beyond restrictions (Hassan *et al.*, 2016). One such tripartite is rhizoremediation, which is practically done by the mixture of two techniques bio-augmentation and phytoremediation. The modern and latest method of mining metals from polluted soil through plant consumption is (phytoextraction), and this can help in observing better yield through another procedure known as phytoremediation (Hamzah *et al.*, 2016). The process of bio-augmentation include the addition of microbes to “support” biological waste treatment for effective reduction of pollution by converting the waste into less harmful compounds (Herrero and Stuckey, 2015). The symbiotic and non-symbiotic associations amongst plants and microbes, which are truly done by PGPR, is one of the source of rhizoremediation. Nowadays, PGPR application through rhizoremediation possess limited research for few species of microbes, for instance *Bacillus* species, and *Pseudomonas aeruginosa*; genetically engineered as *Pseudomonas fluorescens* (Kuiper *et al.*, 2004). Moreover, PGPR is applicable in bioremediators on which the world places a huge demand in eradicating pollution in the form of toxic waste and heavy metals from water and soil environment.

3.2. INDIRECT TOOLS

Indirect mechanisms include the method by which PGPR annul or nullify the deadly effects of phytopathogens on plants by the suppressive substances production which has a potential to rise host's natural resistance (Singh and Jha, 2015). Moreover, this method is referred as a passage which support active growth of plants in environmental stress (abiotic stress) or shield plants from contaminations such as biotic stress (Akhgare *et al.* 2014). The PGPR contribute in this method of releasing of hydrolytic enzymes like (proteases, cellulases, chitinases, etc.), VOCs, EPSs, production of siderophores various antibiotics in response to plant pathogen or disease resistance, induction of systematic resistance against various pathogen and pests, etc. (Nivya, 2015; Gupta *et al.*, 2014).

3.2.1. STRESS CONTROL

The character which is a source of damaging influence on growth of plants is termed as stress (Foyer *et al.*, 2016). Stress of any type causes the rise in the construction of reactive oxygen species (ROS) for instance OH^\cdot , O^{2-} , and H_2O_2 radicals. Extra ROS assembly leads to oxidative stress, that create negative influence on plants with nucleic acids, proteins, membrane lipids and oxidizing photosynthetic pigments. Plants are normally exposed to several ecological stresses and have produced particular response mechanisms (Ramegowda and Senthil-Kumar, 2015). Within the recent ten years, a need for grasping the molecular mechanisms has arisen which has application on biotic and abiotic stress tolerance (Tripathi *et al.*, 2015; Tripathi *et al.*, 2016; Pontigo *et al.*, 2017; Singh *et al.*, 2017; Tripathi *et al.*, 2017). Few of the factors which include PGPR induce stress controlling in plants are discussed [here](#).

(A) NON-BIOLOGICAL STRESS TOLERANCE

Abiotic stress (floods, salinity, drought, extreme temperature and high wind etc.) have an extraordinary damaging effect upon biomass production, therefore a need for survival of essential diet crops specifically up to 70%, is needed which will annul the negative effect on food safety globally. Drought stress reported by dryness, high temperature and salinity, are termed as major abiotic stress which play role in restricting growth and yield of plants (Vejan *et al.*, 2016). Tolerance which is positive response of stress is quantifiable and multigenic naturally, and involves gathering of definite stress metabolites, for instance abscisic acid, glycine-betaine, proline, poly-sugars and include upregulation in the release of non-enzymatic and enzymatic antioxidants, like glutathione, α -tocopherol, glutathione reductase, ascorbic acid, superoxide dismutase (SOD), ascorbate peroxidase (APX), and catalase (CAT) (Agami *et al.*, 2016). Besides these, there are various additional approaches which lessen the intensity of cellular damage produced through water stress and is fruitful in inducing crop tolerance. It involves exogenous application of PGPR on potential osmolytes, for instance trehalose, glycine betaine, and proline, etc., that has attained great responsiveness for decreasing the stress effect. The application of PGPR during abiotic stress controlling in plants has been widely analysed by bacterial strains of *Pseudomonas fluorescens* and *Pseudomonas putida* which is effective in annulling the lethal impact of cadmium pollution upon barley plants through their ability of scavenging soil cadmium ions (Baharlouei *et al.*, 2011). Furthermore, enhanced leaf water status, specifically in abiotic stress and salinity settings, are said to leave proven results relevant to PGPR effects (Ahmad *et al.*, 2013; Naveed *et al.*, 2014). The formation of a relationship amongst drought resistance and PGPR

has been testified in various crops, inclusively wheat, chickpea and soybean (Ngumbi and Kloepper, 2016). Boosted stress tolerance by salinity in okra by ROS-scavenging enzymes and enhanced water-absorbing efficiency, known to be induced through PGPR, has likewise been proven by Habib *et al.*, 2016.

(B) BIOLOGICAL STRESS TOLERANCE

Different pathogens, like viroids, insects, protists, nematodes, viruses, bacteria and fungi lead to definite decrease in crop production (Haggag *et al.*, 2015). Worldwide agricultural yield is countering a substantial loss of nearly 15% primarily because of phytopathogens (Strange and Scott, 2005). Stress lead to decrease in food production and simultaneously supports resistant crops breeding because of great loss economically. Biotic stress impart hostile effects on plants, involving horticultural plant health, natural habitat ecology, ecosystem nutrient cycling, population dynamics and co-evolution (Gusain *et al.*, 2015). Such difficulties could overcome by means of PGPR, for instance *B. subtilis* strain RMPB44, *P. Favisporus* strain BKB30, *Paenibacillus polymyxa* strains B2, B3, B4, *Bacillus amyloliquefaciens* strain HYD-B17, *B. thuringiensis* strain HYDGRFB19, and *B. licheniformis* strain HYTAPB18. Huge resistance to several types of biotic stress is achieved through inoculation of plants with PGPR cultures via seeds or roots soaked overnight (Ngumbi and Kloepper, 2016).

3.2.2. RESISTANCE AGAINST DISEASE ANTIBIOSIS

Microbial antagonists deal with effect of plant pathogens on agrarian fields which has been proposed a substantial substitute for chemical insecticides. PGPR, such as *Pseudomonas* sp and *Bacillus* sp serve a foremost part for pathogenic microbial inhibition by the release of antibiotics. During the recent ten years; PGPR is a source of antibiotics release contrary to various plant pathogens which serve as a phenomenon for biological control (Ulloa-Ogaz *et al.*, 2015). The Record showed that nearly maximum *Pseudomonas* species has the capacity to release antifungal antibiotics (pyocyanin, N-butylbenzene sulfonamide, , butyrolactones, viscosinamide, 2,4-diacetylphloroglucinol, pyoluteorin, pyrrolnitrin, rhamnolipids, cepaciamide A, oomycin A, ecomycins, phenazine-1-carboxamide, phenazines, and phenazine-1-carboxylic acid), antiviral antibiotics (Karalicine), antitumor antibiotics (FR901463 and cepafungins), bacterial antibiotics (pseudomonic acid and azomycin), (Ramadan *et al.*, 2016). *Bacillus* sp. even play a major role for the synthesis of antibacterial and antifungal antibiotics. The source of these antibiotics are both non-ribosomal and ribosomal. The ribosomal source of antibiotics cover sublancin A, subtilintin A, and

subtilosin and the non-ribosomal sources cover bacillaene, difficidin, rhizoctinins, mycobacillin and chlorotetain bacilysin, etc. *Bacillus* sp. also serve as a source of widespread diversity of lipopeptide antibiotics, for instance bacillomycin, iturins, and surfactin, etc. (Wang *et al.*, 2015). The antibiotics are furthermore congregated into volatile and non-volatile compounds. The volatile antibiotics consist of hydrogen cyanide, sulfides, alcohols, ketones, and aldehydes, etc., and the antibiotics which are non-volatile are heterocyclic nitrogenous compounds, phenylpyrrole, polyketides, cyclic lipopeptides, aminopolyols, etc. (Fouzia *et al.*, 2015).

3.2.3. TRIGGERING SYSTEMIC RESISTANCE

Induced systemic resistance (ISR) is termed as a biological condition of enhanced protective potential aroused as a reaction to a specific ecological stimulus. PGPR causes systemic resistance in numerous plants with the effect of various ecological stressors (Prathap and Ranjitha, 2015). Signals are aroused as a protective mechanism stimulated by the vascular system throughout microbial attack that consequently lead to switching on of numerous defence enzymes, like APX, CAT, SOD, lipoxygenase, polyphenol oxidase, β -1, 3glucanase, chitinase, phenylalanine ammonia lyase, and peroxidase along with few proteinase inhibitor. ISR is not specifically applicable upon a certain microbe however it is supportive for plants in combatting various diseases (Kamal *et al.*, 2014). ISR includes signalling of ethylene hormone inside the plant and advantageous in causing host plant's protective responses as an effect of attack of various pathogens of plants. The diverse forms exist in individual bacterial components which are responsible for triggering the release of volatiles, such as acetoin, and 2, 3butanediol, ISR; like lipopolysaccharides, homoserine lactones, 2, 4diacetylphloroglucinol, siderophores, cyclic lipopeptides (Berendsen *et al.*, 2015). While, widespread induction of ISR through PGPR within plants, and their application is regarded as an indispensable source to modernise farming, therefore unpretentious investigation is being carried out by consuming PGPR which lead to a dire demand of modern fundamental tools and procedures usages for sustenance of plants from laboratory to actual field.

3.2.4. APPLICATION OF PROTECTIVE ENZYMES

PGPR plant growth is supported by PGPR by release of metabolites which is a regulator of phytopathogenic agents (Meena *et al.*, 2016). The enzymes like chitinase, ACC- deaminase, and β -1,3-glucanase, which are responsible for breakdown of cell walls and deactivating microbes (Goswami *et al.*, 2016). Generally cell wall of fungi have components consisting of

chitin, and β -1, 4-N-acetyl-glucosamine and hence chitinase-producing bacteria and β -1,3-glucanase-, have potential to control their absolute growth. *Sinorhizobium fredii* KCC5 and *Pseudomonas fluorescens* LPK2 release chitinase and beta-glucanases lead to withering of *Fusarium udum* and *Fusarium oxysporum* (Ramadan *et al.* 2016). *Rhizoctonia solani* and *Phytophthora capsici* are completely inhibited through PGPR because they are considered as the best disastrous crop microbes renowned globally (Islam *et al.*, 2016).

3.2.5. APPLICATION OF VOCs

VOCs which support plant growth since they are released by those strains which show biological control, cause inhibition of microbes such as nematodes, fungi and bacteria, and bring induction of systemic resistance within plants for phytopathogens (Raza *et al.*, 2016a,b). specific species of bacteria belonging from various genera, comprising of *Serratia*, *Stenotrophomonas*, *Arthrobacter*, *Bacillus*, and *Pseudomonas*, have potential to produce VOCs which effect growth of plant. *Bacillus* sp are good producers of acetoin and 2, 3-Butanediol; termed as VOCs for support of growth of plant but cause inhibition of fungal growth (Santoro *et al.*, 2016). It has been testified that plant ISR are produced through bacterial VOCs (Sharifi and Ryu, 2016). The VOCs obtained through PGPR strains indirectly or directly facilitate improved resistance against diseases, improvement in plant biomass and abiotic stress tolerance. The microbes which are responsible for VOC release are 11-decyldocosane, dotriacontane, cyclohexane, 2,6,10-trimethyl, 2-(benzyloxy)ethanamine, tetradecane, benzene, methyl, 1-chlorooctadecane, decane, benzene(1-methylnonadecyl), dodecane, 1-(N-phenylcarbonyl)-2-morpholinocyclohexene. These VOCs are highly demanded in spite of variability existing between identity and quantity of the VOCs released from various species (Kanchiswamy *et al.*, 2015).

4. Upcoming viewpoint

PGPR has been promoting the agricultural yield by means of various processes and mechanisms. Still, several environmental factors influence plant growth through inconsistency in PGPR performance. The environmental factors consist of soil composition or activity of the native microorganism's flora of soil, characteristics of soil, climate of soil, and weather conditions (Gupta *et al.*, 2015). Various types of factors such as abiotic and biotic comprising of herbicides, microbes and weeds etc. which confine PGPR impact causing poor yield in plants. With the arrival of recent techniques and tools for instance Nano-fertilizers, Biosensors and Nanomaterials in the areas of nanotechnology and biotechnology results in

improvement in agricultural yield throughout the latest ten years. Soil serve as the richest constituent containing natural nanoparticles, both of aggregates/agglomerates and primary particles. Nano agriculture is needed to introduce biotechnology, nanotechnology, and other areas of science against agricultural sciences for the purpose of replacement of old agricultural techniques to modern ones leading to food safety for rising population (Subramanian and Tarafdar, 2011). The development of latest Nano devices like (enzyme encapsulation, biosensors,) and nanomaterials like (quantum dots, fullerene derivatives, nanotubes, and nanowires) by the arising of nanotechnology which declare possible narrative use in the area of life sciences and agriculture (Dixshit *et al.*, 2013). The superiority of these materials show reliance on its size in many areas supporting agriculture growth. Plant pathology dependant on nanoparticles is applicable on particular agricultural issues arising during pathogen-plant interactions and offer new techniques for agricultural safety. This consist of timely finding of biological stresses and their control, increasing input for good output like (fruits, flowers and vegetables etc.). Since, PGPR (*Bacillus subtilis*, *Pseudomonas putida*, and *Pseudomonas fluorescens*) treated with silver, gold, and aluminium coated nanoparticles have been testified for the purpose of rise in growth of plant, but also have application in inhibition of the unsafe fungal parasite growth inside rhizosphere, thus performing as possible nano-biofertilizers. The nano bio-fertilizers can be encapsulated through micro-encapsulation and have application in controlling the fertilizer release onto aimed cell in the absence of unintentional loss. Improved holding capacity of useful bacteria with the roots of oil seed rape which is vital for the safety of plants for infection of fungal microbes by Titania nanoparticles was practically proven from Mishra and Kumar, (2009). Percentage of seed germination in various monocots and dicots have also be accepted to be enhanced through pretreatment by ZnO nanoparticles (Mishra and Kumar, 2009). In present situation, the application of nano bio-fertilizers serve a huge prospect to cultivate environmentally friendly compounds which is used as a substitute instead of chemical pesticides (Caraglia *et al.*, 2011). Microencapsulation and Nano encapsulation of nematicides, fungicides and insecticides, are supportive in generating a design which is influential while serving as a pest controller thus inhibiting soil residue. Encapsulated herbicide molecules for instance metal achlor and pentimethalin using polymers like poly alylamine hydrochloride (PAH), and poly styrene sulphonate (PSS) which show lysis in moist condition and therefore definitely be controlled. These encapsulated herbicides have constant production of active ingredients which guarantee productive weed control (Kanimozhi and Chinnamuthu, 2012). Furthermore, these include hydro and thermal strength.

These encapsulated nematicides, fungicides, and insecticides might be supportive in production of formulations implied for pests control. Arrival of smart biosensors for nutrient and contaminants discovery place an enormous effect on precision agriculture that can definitely utilize remote sensing devices, global positioning system (GPS), and computers, for measuring extremely limited ecological conditions, using resources having extreme efficiency and identifying the location and nature of difficulty. Precision farming has been a lengthy demanded objective to decrease input (like herbicide, pesticides, fertilizer, etc.) and make best use of output (like agricultural production) by checking ecological variables and application action directed. Nano scale Zeolites that is referred as naturally existing crystalline aluminum silicates, may even serve major part through enhancing the water retention potential of sandy soil and increase porosity of clay soils (Srilatha, 2011; Subramanian and Tarafdar, 2011; Vacheron *et al.*, 2013; Trivedi and Hemantaranjan, 2014). Bioremediation even has arisen as a budding tool for removing contamination of metal polluted/ environment. The decrease in metal contaminants bioavailability is observed inside rhizosphere (phytostabilization) which is helpful in enhancing plant health, growth and establishment, and can chiefly catalyze growth of plant and definitely its yield (Ma *et al.*, 2011). Production of superior or new strains of PGPR by enhancing above qualities is a probable result of genetic manipulations. These PGPR-biotechnologies can be misused and considered as a low-input, supportable and eco-friendly technique for controlling plant stresses. In present situation Nano based products and techniques have been observed for agronomic development. These techniques are followed in various developed countries such as India, South Korea, Switzerland, USA, Japan, Germany, France, and China where big scale application of such products are still restricted to certain biotechnological foodstuffs like Flavr Savr tomatoes, seedless bananas, Golden rice, BT Brinjal, Cucumber, and BT cotton, etc. and thus main progress is demanded to please the requirements for the growing population.

5. CONCLUSION

Soil and agriculture are two vital components of globe which are necessary for human survival. Undiscriminating misuse of resources has put limitations on food yield which has led mankind to search for alternate for satisfying their requirements for living. PGPR shows an essential part in increasing growth of plant; overcoming the polluted earth consisting of sewage and eutrophied water bodies; and monitoring phosphorous runoff, nitrogen, and pesticide pollution. Yet, extraordinary reliance of mankind on compost and pests killers has increased the intensity of pollution and led to disturbance in ecosystem equilibrium.

Furthermore, they have penetrated the food chain by different means. These deviations can disturb microbe-plant interactions by changing biogeochemical cycles and microbial natural science. Still by implying new tools and practices for PGPR improvement play a vital part for the support of farming by increasing the fertility of soil, keeping a stable nutrient cycling having plant tolerance and rise in yield of agriculture. Advanced research has executed on choosing appropriate rhizosphere microorganisms and involve application upon microbial communities by unifying the research fields material science, chemical engineering, agro biotechnology and biotechnology, and to invent new techniques and tools for bringing revolution in agricultural processes.

REFERENCES:

- Abhilash, P.C., Tripathi, V., Edrisi, S.A., Dubey, R.K., Bakshi, M., Dubey, P.K., et al., 2016. Sustainability of crop production from polluted lands. *Energy Ecol. Environ.* 1 (1), 54–65.
- Agami, R.A., Medani, R.A., Abd El-Mola, I.A., Taha, R.S., 2016. Exogenous application with plant growth promoting rhizobacteria (PGPR) or proline induces stress tolerance in basil plants (*Ocimum basilicum* L.) exposed to water stress. *Int. J. Environ. Agric. Res.* 2 (5), 78–92.
- Agler, M.T., Ruhe, J., Kroll, S., Morhenn, C.M., Kim, S.T., Weigel, D., Kemen, E.M., 2016. Microbial hub Taxa link host and abiotic factors to plant microbiome variation. *PLoS Biol.* 14 (1), 1–31.
- Ahemad, M., Khan, M.S., 2012. Evaluation of plant-growth promoting activities of rhizobacterium *Pseudomonas putida* under herbicide stress. *Ann. Microbiol.* 62, 1531–1540.
- Ahemad, M., Kibret, M., 2014. Mechanisms and applications of plant growth promoting rhizobacteria: current perspective. *J. King Saud Univ. Sci.* 26, 1–20.
- Ahmad, M., Zahir, Z.A., Khalid, M., 2013. Efficacy of *Rhizobium* and *Pseudomonas* strains to improve physiology, ionic balance and quality of mung bean under salt-affected conditions on farmer's fields. *Plant Physiol. Biochem.* 63, 170–176.
- Akhgar, R., Arzanlou, M., Bakker, P.A.H.M., Hamidpour, M., 2014. Characterization of laminocyclopropane-1-carboxylate (ACC) deaminase-containing *Pseudomonas* sp. in the rhizosphere of salt-stressed canola. *Pedosphere* 24, 161–468.

Akhtar, N., Qureshi, M.A., Iqbal, A., Ahmad, M.J., Khan, K.H., 2012. Influence of Azotobacter and IAA on symbiotic performance of Rhizobium and yield parameters of lentil. *J. Agric. Res.* 50, 361–372.

Alexandratos, N., Bruinsma, J., 2003. In: Bruinsma, J. (Ed.), *World Agriculture: Towards 2015/2030 an FAO Perspective*. Earthscan Publications, pp. 1–28.

Anand, K., Kumari, B., Mallick, M.A., 2016. Phosphate solubilizing microbes: an effective and alternative approach as bio-fertilizers. *Int. J. Pharm. Sci.* 8 (2), 37–40.

Araujo, A.S.F., Leite, L.F.C., Santos, V.B., Carneiro, R.F.V., 2009. Soil microbial activity in conventional and organic agricultural system. *Sustainability* 1, 268–276.

Baharlouei, J., Pazira, E., Khavazi, K., Solhi, M., 2011. Evaluation of inoculation of plant growth-promoting rhizobacteria on cadmium uptake by canola and barley. *2nd Int. Conf. Env. Sci. Tech.* 2, 28–32.

Barea, J.M., Pozo, M.J., Azcon, R., Aguilar, C.A., 2005. Microbial co-operation in the rhizosphere. *J. Exp. Bot.* 56, 1761–1778.

Belimov, A.A., Kunakova, A.M., Safronova, V.I., et al., 2004. Employment of rhizobacteria for the inoculation of barley plants cultivated in soil contaminated with lead and cadmium. *Microbiol* 73, 99–106.

Beneduzi, A., Ambrosini, A., Passaglia, L.M.P., 2012. Plant growth-promoting rhizobacteria: their potential as antagonists and biocontrol agents. *Genet. Mol. Biol.* 35 (4), 1044–1051.

Berendsen, R.L., Verk, M.C.V., Stringlis, I.A., Zamioudis, C., Tommassen, J., Pieterse, C.M.J., Bakker, P.A.H.M., 2015. Unearthing the genomes of plant-beneficial *Pseudomonas* model strains WCS358, WCS374 and WCS417. *BMC Genomics* 16, 539.

Bhardwaj, D., Ansari, M.W., Sahoo, R.K., Tuteja, N., 2014. Biofertilizers function as key player in sustainable agriculture by improving soil fertility, plant tolerance and crop productivity. *Microbial. Cell Fact.* 13 (66), 1–10.

Bhattacharyya, P.N., Jha, D.K., 2012. Plant growth-promoting rhizobacteria (PGPR): emergence in agriculture. *World J. Microbial. Biotechnol.* 28, 1327–1350.

Caraglia, M., Rosa, G.D., Abbruzzese, A., Leonetti, C., 2011. Nanotechnologies: new opportunities for old drugs. the case of amino-bisphosphonates. *Nanomedic. Biotherapeu Dis.* 1, 103e.

Compant, S., Saikkonen, K., Mitter, B., Campisano, A., Blanco, J.M., 2016. Soil, plants and endophytes. *Plant Soil* 405, 1–11.

Damam, M., Kaloori, K., Gaddam, B., Kausar, R., 2016. Plant growth promoting substances (phytohormones) produced by rhizobacterial strains isolated from the rhizosphere of medicinal plants. *Int. J. Pharm. Sci. Rev.* 37 (1), 130–136.

Damir, O., Mladen, P., Bozidar, S., Srnan, N., 2011. Cultivation of the bacterium *Azotobacter chroococcum* for preparation of biofertilizers. *Afr. J. Biotechnol.* 10, 3104–3111.

Dixshit, A., Shukla, S.K., Mishra, R.K., 2013. Exploring Nanomaterials with PGPR in Current Agriculture Scenario PGPR with Special Reference to Nanomaterials. Lab Lambert Academic Publication, Germany 51 pp.

Egamberdieva, D., Lugtenberg, B., 2014. Use of plant growth-promoting rhizobacteria to alleviate salinity stress in plants. PGPR to alleviate salinity stress on plant growth. In: Miransari, M. (Ed.), *Use of Microbes for the Alleviation of Soil Stresses*. Springer, New York, pp. 73–96.

Fouzia, A., Allaoua, S., Hafsa, C., Mostefa, G., 2015. Plant growth promoting and antagonistic traits of indigenous fluorescent *Pseudomonas* spp. Isolated from wheat rhizosphere and a thalamus endosphere. *Eur. Sci. J.* 11, 129–148.

Foyer, C.H., Rasool, B., Davey, J.W., Hancock, R.D., 2016. Cross-tolerance to biotic and abiotic stresses in plants: a focus on resistance to aphid infestation. *J. Exp. Bot.* 7, 2025–2037.

Gabriela, F., Casanovas, E.M., Quillehauquy, V., Yommi, A.K., Goni, M.G., Roura, S.I., Barassi, C.A., 2015. *Azospirillum* inoculation effects on growth, product quality and storage life of lettuce plants grown under salt stress. *Scientia Horticul.* 195, 154–162.

Garcia, F.P., Menendez, E., Rivas, R., 2015. Role of bacterial bio fertilizers in agriculture and forestry. *AIMS Bioeng.* 2, 183–205.

Glick, B.R., 2012. Plant growth-promoting bacteria: mechanisms and applications. *Scientifica* 1–15.

Gonzalez, A.J., Larraburu, E.E., Llorente, B.E., 2015. Azospirillumbrasilense increased salt tolerance of Jojoba during in vitro rooting. *Ind. Crops Products* 76, 41–48.

Goswami, D., Pithwa, S., Dhandhukia, P., Thakker, J.N., 2014. Delineating Kocuriaturfanensis 2M4 as a credible PGPR: A novel IAA producing bacteria isolated from saline desert. *J. Plant Interact.* 9, 566–576.

Goswami, D., Thakker, J.N., Dhandhukia, P.C., 2016. Portraying mechanics of plant growth promoting rhizobacteria (PGPR): a review. *Cogent Food Agric.* 2, 1–19.

Gray, E.J., Smith, D.L., 2005. Intracellular and extracellular PGPR: Commonalities and distinctions in the plant-bacterium signaling processes. *Soil Biol. Biochem.* 37, 395–412.

Gupta, S., Meena, M.K., Datta, S., 2014. Isolation, characterization of plant growth promoting bacteria from the plant Chlorophytumborivilianum and in-vitro screening for activity of nitrogen fixation, phosphate solubilization and IAA production. *Int. J. Curr. Microbial. Appl. Sci.* 3, 1082–1090.

Gupta, G., Parihar, S.S., Ahirwar, N.K., Snehi, S.K., Singh, V., 2015. Plant growth promoting Rhizobacteria (PGPR): Current and future prospects for development of sustainable agriculture. *J. Microbiol. Biochem.* 7, 96–102.

Gusain, Y.S., Singh, U.S., Sharma, A.K., 2015. Bacterial mediated amelioration of drought stress in drought tolerant and susceptible cultivars of rice (*Oryzasativa* L.). *Afr. J. Biotechnol.* 14, 764–773.

Habib, S.H., Kausar, H., Saud, H., 2016. Plant growth promoting rhizobacteria enhance salinitystresstoleranceinOkrathrough ROS-Scavenging enzymes. *Bio.Med.Res.Int.* 1–10.

Haggag, W.M., Abouziena, H.F., Abd-El-Kreem, F., Habbasha, S., 2015. Agriculture biotechnology for management of multiple biotic and abiotic environmental stress in crops. *J. Chem. Pharm.* 7 (10), 882–889.

Hamzah, A., Hapsari, R.I., Wisnubroto, E.I., 2016. Phytoremediation of Cadmium-contaminated agricultural land using indigenous plants. *Int. J. Environ. Agric. Res.* 2 (1), 8–14.

Hassan, I., Mohamed elhassan, E., Yanful, E.K., Yuan, Z., 2016. A Review Article: electrokinetic bioremediation current knowledge and new prospects. *Adv. Microbiol.* 6, 57–72.

Herrero, M., Stuckey, D.C., 2015. Bio-augmentation and its application in waste water treatment: a review. *Chemosphere* 140, 119–128.

Islam, S., Akanda, A.M., Prova, A., Islam, Md. T., Hossain, Md, Md., 2016. Isolation and identification of plant growth promoting rhizobacteria from cucumber rhizosphere and their effect on plant growth promotion and disease suppression. *Front. Microbiol.* 6 (1360), 1–12.

Jahanian, A., Chaichi, M.R., Rezaei, K., Rezayazdi, K., Khavazi, K., 2012. The effect of plant growth promoting rhizobacteria (PGPR) on germination and primary growth of artichoke (*Cynarascolymus*). *Int. J. Agric. Crop Sci.* 4, 923–929.

Jung, H., Janelle, K., McCouch, S., 2013. Getting to the roots of it: genetic and hormonal control of root architecture. *Front. Plant Sci.* 4 (186), 1–32.

Kamal, R., Gusain, Y.S., Kumar, V., 2014. Interaction and symbiosis of fungi, Actinomycetes and plant growth promoting rhizobacteria with plants: strategies for the improvement of plants health and defense system. *Int. J. Curr. Microbial. Appl. Sci.* 3 (7), 564–585.

Kamkar, B., 2016. Sustainable development principles for agricultural activities. *Adv. Plant Agric. Res.* 3 (5), 1–2.

Kanchiswamy, C.N., Malnoy, M., Maffei, M.E., 2015. Chemical diversity of microbial volatiles and their potential for plant growth and productivity. *Front. Plant Sci.* 6, 151.

Kang, B.G., Kim, W.T., Yun, H.S., Chang, S.C., 2010. Use of plant growth-promoting rhizobacteria to control stress responses of plant roots. *Plant Biotechnol. Rep.* 4, 179–183.

Kanimozhi, V., Chinnamuthu, C.R., 2012. Engineering Core/hallow shell nanomaterials to load herbicide active ingredient for controlled release. *Res. J. Nanosci. Nanotechnol.* 2, 58–69.

Khan, Z., Rho, H., Firrincieli, A., Hung, H., Luna, V., Masciarelli, O., Kim, S.H., Doty, S.L., 2016. Growth enhancement and drought tolerance of hybrid poplar upon inoculation with endophyte consortia. *Curr. Plant Biol.* 1–10.

Kuan, K.B., Othman, R., Rahim, K.A., Shamsuddin, Z.H., 2016. Plant growth-promoting rhizobacteria inoculation to enhance vegetative growth, Nitrogen fixation and nitrogen remobilisation of Maize under Greenhouse conditions. *PLoS One* 11 (3), 1–19.

Kuiper, I., Lagendijk, E.L., Bloemberg, G.V., Lugtenberg, B.J.J., 2004. Rhizoremediation: a beneficial plant-microbe interaction. *Mol. Plant Microbe Inter.* 7 (1), 6–15.

Kumar, P., Dubey, R.C., 2012. Plant Growth Promoting Rhizobacteria for biocontrol of phytopathogens and yield enhancement of *Phaseolus vulgaris*. *J. Curr. Perspect. App. Microbiol.* 1, 6–38.

Kumar, A.D.S., Vidhya, A.K., Ragunathan, R., Johney, J., 2015. Production and purification and characterization of streptokinase using *Bacillus licheniformis* under solid state fermentation. *J. Glob. Biosci.* 4 (7), 2703–2712.

Kumar, A., 2016. Phosphate solubilizing bacteria in agriculture biotechnology: diversity, mechanism and their role in plant growth and crop yield. *Int. J. Adv. Res.* 4 (4), 116–124.

Kundan, R., Pant, G., Jado, N., Agrawal, P.K., 2015. Plant growth promoting rhizobacteria: mechanism and current prospective. *J. Fertilizers Pesticides* 6, 2.

Lehmann, J., Kleber, M., 2015. The contentious nature of soil organic matter. *Nature* 528, 60–68.

Liu, D., Lian, B., Dong, H., 2012. Isolation of *Paenibacillus* sp. and assessment of its potential for enhancing mineral weathering. *J. Geomicrobiol.* 29, 413–421.

Liu, W., Wang, Q., Hou, J., Tu, C., Luo, Y., Christie, P., 2016. Whole genome analysis of halotolerant and alkalotolerant plant growth-promoting rhizobacterium *Klebsiella* sp. D5A. *Sci. Rep.* 6, 26710.

Ma, Y., Rajkumar, M., Freitas, H., 2009. Inoculation of plant growth promoting bacterium *Achromobacter xylosoxidans* strain Ax10 for the improvement of copper phytoextraction by *Brassica juncea*. *J. Environ. Manag.* 90, 831–837.

Ma, Y., Prasad, M.N.V., Rajkuma, M., Freitas, H., 2011. Plant growth promoting rhizobacteria and endophytes accelerate phytoremediation of metalliferous soils. *Biotechnol. Adv.* 29, 248–258.

Mahmood, S., Daur, I., Al-Solaimani, S.G., Ahmad, S., Madkour, M.H., Yasir, M., Hirt, H., Ali, S., Ali, Z., 2016. Plant growth promoting rhizobacteria and silicon synergistically enhance salinity tolerance of mung bean. *Front. Plant Sci.* 7, 1–14.

Meena, M.K., Gupta, S., Datta, S., 2016. Antifungal Potential of PGPR, their growth promoting activity on seed germination and seedling growth of winter wheat and genetic variability among bacterial isolates. *Int. J. Curr. Microbial. Appl. Sci.* 5 (1), 235–243.

Mishra, V.K., Kumar, A., 2009. Impact of metal nanoparticles on the plant growth promoting rhizobacteria. *Digest J. Nanomat. Biostruct.* 4 (3), 587–592.

Montano, F.P., Villegas, C.A., Bellogia, R.A., Cerro, P.D., Espuny, M.R., Guerrero, I.J., et al., 2014. Plant growth promotion in cereals and leguminous agricultural important plants from microorganisms capacities to crop production. *Microbiol. Res.* 169 (5–6), 325–336.

Murai, N., 2014. Review: plant growth hormone Cytokinins control the crop seed yield. *Am. J. Plant Sci.* 5, 2178–2187.

Mus, F., Crook, M.B., Garcia, K., Costas, A.G., Geddes, B.A., Kouri, E.D., Paramasivan, P., et al., 2016. Symbiotic Nitrogen fixation and the challenges to its extension to nonlegumes. *Appl. Environ. Microbiol.* 82 (13), 3698–3710.

Narozna, D., Pudełko, K., Kroliczak, J., Golinska, B., Sugawara, M., Cezary, J., et al., 2014. Survival and Competitiveness of Bradyrhizobium japonicum Strains 20 Years after introduction into field locations in Poland. *Appl. Environ. Microbiol.* 81, 5551–5559.

Naveed, M., Hussain, M.B., Zahir, Z.A., Mitter, B., Sessitsch, A., 2014. Drought stress amelioration in wheat through inoculation with Burkholderia phytofirmans strain PsJN. *Plant Growth Regul.* 73, 121–131.

Ngumbi, E., Kloepper, J., 2016. Bacterial-mediated drought tolerance: current and future prospects. *Appl. Soil Ecol.* 105, 109–125.

Nivya, R.M., 2015. A study on plant growth promoting activity of the Endophytic bacteria isolated from the root nodules of Mimosa pudica Plant. *Int. J. Innov. Res. Sci. Er. Technol.* 4, 6959–6968.

Orlandini, V., Emiliani, G., Fondi, M., Maida, E., Perrin, E., Fani, R., 2014. Network Analysis of Plasmidomes: The Azospirillum Brasilense Sp245 Case. Hindawi Publishing Corporation, pp. 1–14.

Oteino, N., Lally, R.D., Kiwanuka, S., Lloyd, A., Ryan, D., Germaine, K.J., Dowling, D.N., 2015. Plant growth promotion induced by phosphate solubilizing endophytic Pseudomonas isolates. *Front. Microbiol.* 6, 745.

Oyedele, Ogunbanwo, A.O., Samuel, T., 2014. Antifungal activities of *Bacillus subtilis* isolated from some condiments and soil. *Afr. J. Microbiol. Res.* 8 (18), 1841–1849.

Paredes, S.H., Lebeis, S.L., 2016. Giving back to the community: microbial mechanisms of plant–soil interactions. *Funct. Ecol.* 30 (7), 1–10.

Parmar, P., Sindhu, S.S., 2013. Potassium solubilisation by Rhizosphere Bacteria: influence of nutritional and environmental conditions. *J. Microbiol. Res.* 3, 25–31.

Passari, A.K., Chandra, P., Zothanpuia, Mishra, V.K., Leo, V.V., Gupta, V.K., Kumar, B., Singh, B.P., 2016. Detection of biosynthetic gene and phytohormone production by endophytic actinobacteria associated with *Solanum lycopersicum* and their plant growth-promoting effect. *Res. Microbiol.* 167 (8), 692–705.

Paustian, K., Lehmann, J., Ogle, S., Reay, D., Robertson, P.G., Smith, P., 2016. Climate-smart soils. *Nature* 532, 49–57.

Pawar, S.T., Bhosale, A.A., Gawade, T.B., Nale, T.R., 2016. Isolation, screening and optimization of exo-polysaccharide producing bacterium from saline soil. *J. Microbiol. Biotechnol. Res.* 3 (3), 24–31.

Perez, Y.M., Charest, C., Dalpe, Y., Seguin, S., Wang, X., Khanizadeh, S., 2016. Effect of inoculation with arbuscular mycorrhizal fungus on selected spring wheat lines. *Sustain. Agric. Res.* 5 (4), 24–29.

Pontigo, S., Godoy, K., Jiménez, H., Gutierrez-Moraga, A., Mora, M.D.L.L., Cartes, P., 2017. Silicon-mediated alleviation of aluminum toxicity by modulation of Al/Si uptake and antioxidant performance in ryegrass plants. *Front. Plant Sci.* 8, 642.

Prathap, M., Ranjitha, K.B.D., 2015. A Critical review on plant growth promoting rhizobacteria. *J. Plant Pathol. Microbiol.* 6 (4), 1–4.

Ramadan, E.M., AbdelHafez, A.A., Hassan, E.A., Saber, F.M., 2016. Plant growth promoting rhizobacteria and their potential for biocontrol of phytopathogens. *Afr. J. Microbiol. Res.* 10, 486–504.

Ramegowda, V., Senthil-Kumar, M., 2015. The interactive effects of simultaneous biotic and abiotic stresses on plants: mechanistic understanding from drought and pathogen combination. *J. Plant Physiol.* 176, 47–54.

Rathore, P., 2015. A review on approaches to develop plant growth promoting rhizobacteria. *Int. J. Recent Sci. Res.* 5 (2), 403–407.

Raza, W., Ling, N., Yang, L., Huang, Q., Shen, Q., 2016a. Response of tomato wilt pathogen *Ralstoniasolanacearum* to the volatile organic compounds produced by a biocontrol strain *Bacillus amyloliquefaciens* SQR-9. *Sci. Rep.* 6, 24856.

Raza, W., Yousaf, S., Rajer, F.U., 2016b. Plant growth promoting activity of volatile organic compounds produced by Bio-control strains. *Sci. Lett.* 4 (1), 40–43.

Reed, S.C., Cleveland, C.C., Townsend, A.R., 2011. Functional ecology of free-living nitrogen fixation: a contemporary perspective. *Annu. Rev. Ecol. Environ. Syst.* 42, 489–512.

Richa, S., Subhash, C., Singh, A., 2013. Isolation of microorganism from soil contaminated with degraded paper in Jharna village. *J. Soil Sci. Environ. Manag.* 4 (2), 23-27.

Rosenberg, E., Rosenberg, I.Z., 2016. Microbes drive evolution of animals and plants: the Hologenome concept. *Microbial. Biol.* 7 (2), 1–8.

Ross, P.J., Holland, S.M., Gill, V.J., Gallin, J.I., 1995. Severe Burkholderia (*Pseudomonas*) *gladioli* infection in chronic granulomatous disease: report of two successfully treated cases. *Clin. Infect. Dis.* 21 (5), 1291–1293.

Rout, M.E., Callaway, R.M., 2012. Interactions between exotic invasive plants and soil microbes in the rhizosphere suggest that everything is not everywhere. *Ann. Bot.* 110, 213–222.

Saha, M., Sarkar, S., Sarkar, B., Sharma, B.K., Bhattacharjee, S., Tribedi, P., 2016. Microbial siderophores and their potential applications: a review. *Environ. Sci. Pollut. Res.* 23 (5), 3984–3999.

Sanders, J.W., Martin, J.W., Hooke, M., Hooke, J., 2000. *Methylobacterium mesophilicum* infection: case report and literature review of an unusual opportunistic pathogen. *Clin. Infect. Dis.* 30 (6), 936–938.

Sanlibaba, P., Cakmak, G.A., 2016. Exo-polysaccharides production by lactic acid bacteria. *Appl. Microbiol.* 2 (2), 1–5.

Santoro, M.V., Bogino, P.C., Nocelli, N., Cappellari, L.R., Giordano, W.F., Banchio, E., 2016. Analysis of plant growth promoting effects of Fluorescent *Pseudomonas* strains isolated

from *Menthapiperita* Rhizosphere and effects of their volatile organic compounds on essential oil composition. *Front. Microbiol.* 7 (1085), 1–17.

Setiawati, T.C., Mutmainnah, L., 2016. Solubilization of Potassium containing mineral by microorganisms from sugarcane Rhizosphere. *Agri. Sci. Procedia* 9, 108–117.

Sharifi,R., Ryu, C.M., 2016. Arebacterial volatile compounds poisonous odors to afungal pathogen *Botrytis cinerea*, Alarm signals to *Arabidopsis* seedlings for eliciting induced resistance, or both? *Front. Microbiol.* 7 (196), 1–10.

Sharma, A., Johri, B.N., Sharma, A.K., Glick, B.R., 2013. Plant growth-promoting bacterium *Pseudomonas* sp. strain GRP3 influences iron acquisition in mung bean (*Vignaradiata*L. Wilzeck). *Soil Biol. Biochem.* 35, 887–894.

Shrestha, J., 2016. A review on sustainable agricultural intensification in Nepal. *Int. J. Bus. Soc. Sci. Res.* 4 (3), 152–156.

Shridhar,B.S.,2012.Review: nitrogen fixingmicroorganisms.*Int.J.Microbial.Res.*3(1), 46–52.

Singh, R.P., Jha, P.N., 2015. Molecular identification and characterization of rhizospheric bacteria for plant growth promoting ability. *Int. J. Curr. Biotechnol.* 3, 12–18.

Singh, S., Tripathi, D.K., Singh, S., Sharma, S., Dubey, N.K., Chauhan, D.K., Vaculík, M., 2017. Toxicity of aluminium on various levels of plant cells and organism: a review. *Environ. Exp. Bot.* 137, 177–193.

Skoog, F., Miller, C.O., 1957. Chemical regulation of growth and organ formation in plant tissue cultures in vitro. *Symp. Soc. Exp. Biol.* 11, 118–131.

Srilatha, B., 2011. Nanotechnology in agriculture. *J. Nanomed. Nanotechnol.* 2, 123.

Stefanescu, I.A., 2015. Bioaccumulation of heavy metals by *Bacillus megaterium*fromphosphogypsum waste. *Sci. Study Res.* 16 (1), 093–097.

Strange, R.N., Scott, P.R., 2005. Plant disease: a threat to global food security. *Ann. Rev. Phytopatho* 43, 1–660.

Subramanian, K.S., Tarafdar, J.C., 2011. Prospects of nanotechnology in Indian farming. *Indian J. Agri. Sci.* 81, 887–893.

Suhag, M., 2016. Potential of biofertilizers to replace chemical fertilizers. *Int. Adv. Res. J. Sci. Eng. Technol.* 3 (5), 163–167.

Sureshababu, K., Amaresan, N., Kumar, K., 2016. Amazing multiple function properties of plant growth promoting rhizobacteria in the rhizosphere soil. *Int. J. Curr. Microbiol. Appl. Sci* 5 (2), 661–683.

Tripathi, D.K., Singh, V.P., Kumar, D., Chauhan, D.K., 2012. Impact of exogenous silicon addition on chromium uptake, growth, mineral elements, oxidative stress, antioxidant capacity, and leaf and root structures in rice seedlings exposed to hexavalent chromium. *Acta Physiol. Plant.* 34, 279–289.

Tripathi, D.K., Singh, V.P., Prasad, S.M., Chauhan, D.K., Dubey, N.K., Rai, A.K., 2015. Silicon-mediated alleviation of Cr (VI) toxicity in wheat seedlings as evidenced by chlorophyll fluorescence, laser induced breakdown spectroscopy and anatomical changes. *Ecotoxicol. Environ. Saf.* 113, 133–144.

Tripathi, D.K., Singh, S., Singh, V.P., Prasad, S.M., Chauhan, D.K., Dubey, N.K., 2016. Silicon nanoparticles more efficiently alleviate arsenate toxicity than silicon in maize cultivar and hybrid differing in arsenate tolerance. *Front. Environ. Sci.* 4, 46.

Tripathi, D.K., Singh, S., Singh, V.P., Prasad, S.M., Dubey, N.K., Chauhan, D.K., 2017. Silicon nanoparticles more effectively alleviated UV-B stress than silicon in wheat (*Triticumaestivum*) seedlings. *Plant Physiol. Biochem.* 110, 70–81.

Trivedi, A.K., Hemantaranjan, A., 2014. Physiological and molecular strategies for crop improvement under changing environment. *Adv. Plant Physiolol.* 15, 1–90.

Tscharntke, T., Clough, Y., Wanger, T.C., Jackson, L., Motzke, I., Perfecto, I., Vandermeer, J., Whitbread, A., 2012. Global food security: biodiversity conservation and the future of agricultural intensification. *Bio Cons* 151, 53–59.

Ubertino, S., Mundler, P., Tamini, L.D., 2016. The adoption of sustainable management practices by Mexican coffee producers. *Sustain. Agric. Res* 5 (4), 1–12.

Ulloa-Ogaz, A.L., Munoz-Castellanos, L.N., Nevarez-Moorillon, G.V., 2015. Biocontrol of phytopathogens: Antibiotic production as mechanism of control, the battle against microbial pathogens. In: In: Mendez Vilas, A. (Ed.), *Basic Science, Technological advance and educational programs* 1. pp. 305–309.

Uqab, B., Mudasir, S., Nazir, R., 2016. Review on bioremediation of pesticides. *J. Biorem. Biodegrad.* 7 (3), 1–5.

Vacheron, J., Desbrosses, G., Bouffaud, M.L., Touraine, B., Moëgne-Loccoz, Y., Muller, D., Legendre, L., Wisniewski-Dyé, F., Combaret, C.P., 2013. Plant growth promoting rhizobacteria and root system functioning. *Front. Plant Sci.* 4 (356), 1–19.

Vejan, P., Abdullah, R., Khadiran, T., Ismail, S., Boyce, A.N., 2016. Role of plant growth promoting Rhizobacteria in agricultural sustainability- a review. *Molecules* 21 (573), 1–17.

Viveros, O.M., Jorquera, M.A., Crowley, D.E., Gajardo, G., Mora, M.L., 2010. Mechanisms and practical considerations involved in plant growth promotion by rhizobacteria. *J. Soil Sci. Plant Nutr.* 10, 293–319.

Wang, X., Mavrodi, D.V., Ke, L., Mavrodi, O.V., Yang, M., Thomashow, L.S., Zheng, N., Weller, D.M., Zhang, J., 2015. Biocontrol and plant growth-promoting activity of rhizobacteria from Chinese fields with contaminated soils. *Microbial. Biotechnol.* 8, 404–418.

Xie, J., Shi, H., Du, Z., Wang, T., Liu, X., Chen, S., 2016. Comparative genomic and functional analysis reveals conservation of plant growth promoting traits in *Paenibacillus polymyxa* and its closely related species. *Sci. Rep.* 6, 21329.

Xu, G., Fan, X., Miller, A.J., 2012. Plant Nitrogen assimilation and use efficiency. *Ann. Rev. Plant Biol.* 63, 153–182.

Zakry, F.A.A., Shamsuddin, Z.H., Khairuddin, A.R., Zakaria, Z.Z., Anuar, A.R., 2012. Inoculation of *Bacillus sphaericus* UPMB-10 to young oil palm and measurement of its uptake of fixed nitrogen using the N isotope dilution technique. *Microbial. Environ.* 27, 257–26.