

for high-thickness epigenome-wide planning of DNA methylation. They saw several examples of epigenetic methylation protected via tissues—the CpGs districts identified as clever biomarkers of HF—and used this innovation to find epigenetic defenselessness and new biomarkers connected with HF and heart failure (Meder *et al.*, 2017; Rau and Vondriska, 2017). In HF patients' blood leukocytes, differently methylation DNA areas were also identified (Li *et al.*, 2017a). The discovery of unusual DNA methylation in the declaration of lymphocyte antigen 75 (LY75) and adenosine receptor A2A (ADORA2A) mRNA in idiopathic widened cardiomyopathy patients was linked to significant variations in the representation of lymphocyte antigen 75 (LY75) and adenosine receptor A2A (ADORA2A) mRNA (Haas *et al.*, 2013). In neurotic and solid hearts, genome-wide guides of DNA methylation and increase of histone three lysine-36 trimethylations (H3K36me3) were also examined. Advertiser CpG islands, quality, intragenic CpG islands, and H3K36me3-rich regions of the genome all showed differences in DNA methylation. Advertisers of elevated traits had changed their DNA methylation, but not those of downregulated qualities. A large number of DUX4 records were linked to differences in DNA methylation and H3K36me3 augmentation. Although more research is needed, there is evidence that the epigenome may limit the declaration of essential features for the improvement of cardiomyopathies (Movassagh *et al.*, 2011). In addition, there is an altered methylation design in the administrative areas of cardiovascular improvement qualities, such as T-box protein 5 (TBX5), heart and neural peak subsidiaries communicated 1 (HAND1), and NK2 homeobox 5 (NKX2.5) in patients with expanding cardiomyopathy (Jo *et al.*, 2016). Koczor *et al.* (2013) examined distinct methylation patterns in patients with enlarged cardiomyopathy, characterized by congestive heart failure (HF). A computational analysis revealed that many great advertisers are differentially methylated (AURKB, BTNL9, CLDN5, and TK1). This study provides valuable information on DNA methylation and adjusted articulation in enlarged cardiomyopathy, which will aid treatment (Koczor *et al.*, 2013).

Furthermore, in the murine model of pressure overburden, epigenetic alterations have been postulated to have a vital role in HF migration. The researchers discovered a drop in sarcoplasmic reticulum Ca ATPase (Atp2a2) levels as well as a compulsory enlistment of -myosin-weighty chain (Myh7) mRNA. Following two months of cross-over aortic tightness, they discovered H3K4me2, H3K9me2, H3K27me3, and H3K36me2, as well as a reduction in the lysine-explicit demethylase KDM2A. (Angrisano *et al.*, 2014). Atp2a2 is a factor in heart capacity, and its decreased mobility is a common feature in HF. Gorski *et al.* (2019) investigated the significance of lysine acetylation in the function of Atp2a2 in HF patients. They discovered

that SIRT1 and Cap p300 regulate acetylation at lysine 492 and significantly reduce gene activity (Gorski *et al.*, 2019). Combining all of this data would be critical in identifying anticipated biomarkers and new epigenetic medicines for HF treatment.

Surprisingly, reactivation of the fetal gene program in HF has been linked to epigenetic remodeling in the preliminary natriuretic peptide (ANP) and BNP promoters. HDAC4, which is sent out from the core, the nucleus, was upregulated in HF patients but not in response to an increase in histone acetylation.

Epigenetic Biomarkers: Limitations and Future Prospects

Epigenetic tools such as DNA methylation and histone changes have been identified as sources of potential biomarkers useful in therapeutic practice. Nonetheless, different epigenetic pathways lead to different CVDs, and different CVDs are regulated by the same epigenetic system, the great majority of which is still under investigation. In hypertension patients, hypermethylation of H3K79 (Rodriguez-Iturbe, 2006; Duarte *et al.*, 2012) and ACE2 advertiser (Fan and al., 2017) has been shown. In addition, both mouse models of hypertension (Pojoga *et al.*, 2011) and HF had hypermethylated H3K4 and H3K9 (Angrisano *et al.*, 2014). This makes selecting and implementing some biomarkers for a given CVD difficult. Another potential concern is the type of the samples, particularly those obtained from pathology division assortments. These specimens are usually preserved in formaldehyde and paraffin, which debase DNA significantly. The length of fixation and storage determines the size, (Kristensen *et al.*, 2009). As a result, a thorough examination of DNA's nature is required. However, in older samples, DNA methylation analysis can be done efficiently using polymerase chain reaction (PCR) techniques with small amplicons (Tournier *et al.*, 2012; Wong *et al.*, 2014). Change the convention with caution in certain situations. Consider that frozen and paraffin-saved samples may yield different results, and they should not be considered without careful consideration (Garca-Giménez *et al.*, 2017).

It's critical to conduct studies with many collaborators in various independent research institutions, all using the same trial design, test arrangement, philosophy, and disease information. Before approving the outcomes of a more extensive sample study, small tolerant partners should be examined as pilot concentrates. The localization approach should be normalized for clinical use, and clinical preliminaries should be randomized and prepared. Compare and contrast the new biomarkers with the traditional biomarkers to determine their usefulness. Each biomarker's affectability and explicitness for a given condition must also be chosen (Engelhardt, 2012; Garca-Giménez *et al.*, 2017). The luminometric methylation measure

and the methylation examination of CpG islands in repeatable components (LINE-1) are widely used techniques for locating DNA methylation. Even though the estimations obtained using the two procedures have a clear relationship, the correlation isn't recommended because a reliable inclination between the outcomes has been exhibited (Knothe *et al.*, 2016). Surprisingly, a significant multicenter investigation comparing possible and routine clinical use of DNA methylation measurements has been conducted. According to the authors, there is a considerable deal of agreement between DNA methylation tests, which may be carried out in a wide range of approval scenarios, the development of new biomarkers, and clinical diagnostics (Outline Consortium, 2016).

Conclusion

Epigenetics and its dynamic cross-talk with inherited traits have received a lot of attention in recent years. Providing a tailored epigenetic example can provide a wealth of information on epigenetic machinery that can be used to customize CVD diagnosis and treatment strategies. DNA methylation, which is regulated by DNA methyltransferases, is often linked to transcriptional restriction, affecting gene articulation by altering the DNA promoter's accessibility to RNA polymerase and consequently gene transcription.

Recent advancements in innovation and data analysis have made it possible to create point-by-point epigenetic maps, which could be used as another tool in clinical practice to assess cardiovascular risk and risk drivers. Epigenetic data can also aid in the prediction of specific drug effects. Crucially, DNA methylation is gaining traction among mainstream academics as a tool for predicting and predicting CVDs. Nonetheless, because of flaws in explicit analytic biomarkers, verification of the current findings is required, with many exploratory communities and large sample size. This will be completed in its entirety.

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REFERENCES

Adachi T., Nakanishi M., Otsuka Y., Nishimura K., Hirokawa G., Goto Y., et al. (2010). Plasma microRNA 499 as a biomarker of acute myocardial infarction. *Clin. Chem.* 56, 1183–1185. 10.1373/clinchem.2010.144121.

Ai J., Zhang R., Li Y., Pu J., Lu Y., Jiao J., et al. (2010). Circulating microRNA-1 as a potential novel biomarker for acute myocardial infarction. *Biochem. Biophys. Res. Commun.* 391, 73–77. 10.1016/j.bbrc.2009.11.005.

Akat K. M., Moore-McGriff D., Morozov P., Brown M., Gogakos T., Correa Da Rosa J., et al. (2014). Comparative RNAsequencing analysis of myocardial and circulating small RNAs in human heart failure and their utility as biomarkers. *Proc. Natl. Acad. Sci. U. S. A* 111, 11151–11156. 10.1073/pnas.1401724111.

Akhtar M. M., Micolucci L., Islam M. S., Olivieri F., Procopio A. D. (2016). Bioinformatic tools for microRNA dissection. *Nucleic Acids Res.* 44, 24–44. 10.1093/nar/gkv1221.

Alavi-Moghaddam M., Chehrazi M., Alipoor S. D., Mohammadi M., Baratloo A., Mahjoub M. P., et al. (2018). A preliminary study of microRNA-208b after acute myocardial infarction: impact on 6-month survival. *Dis. Markers* 2018, 2410451–7. 10.1155/2018/2410451.

Alikhani-Koopaei R., Fouladkou F., Frey F. J., Frey B. M. (2004). Epigenetic regulation of 11 beta-hydroxysteroid dehydrogenase type 2 expression. *J. Clin. Investig.* 114, 1146–1157. 10.1172/JCI21647.

Angrisano T., Schiattarella G. G., Keller S., Pironti G., Florio E., Magliulo F., et al. (2014). Epigenetic switch at *atp2a2* and *myh7* gene promoters in pressure overload-induced heart failure. *PLoS One* 9, e106024. 10.1371/journal.pone.0106024.

Authors/Task Force members. Windecker S., Kolh P., Alfonso F., Collet J.-P., Cremer J., et al. (2014). 2014 ESC/EACTS guidelines on myocardial revascularization: the task force on myocardial revascularization of the European Society of Cardiology (ESC) and the European Association for Cardio-Thoracic Surgery (EACTS) developed with the special contribution of the European Association of Percutaneous Cardiovascular Interventions (EAPCI). *Eur. Heart J.* 35, 2541–2619. 10.1093/eurheartj/ehu278

Babuín L., Jaffe A. S. (2005). Troponin: the biomarker of choice for the detection of cardiac injury. *CMAJ* 173, 1191–1202. 10.1503/cmaj/051291

Baccarelli A., Rienstra M., Benjamin E. J. (2010). Cardiovascular epigenetics: basic concepts and results from animal and human studies. *Circ. Cardiovasc. Genet.* 3, 567–573. 10.1161/CIRCGENETICS.110.958744

Baptista R., Marques C., Catarino S., Enguita F. J., Costa M. C., Matafome P., et al. (2018). MicroRNA-424(322) as a new marker of disease progression in pulmonary arterial hypertension and its role in right ventricular hypertrophy by targeting SMURF1. *Cardiovasc. Res.* 114, 53–64. 10.1093/cvr/cvx187

Bartel D. P. (2009). MicroRNAs: target recognition and regulatory functions. *Cell* 136, 215–233. 10.1016/j.cell.2009.01.002

Bauters C., Kumarswamy R., Holzmann A., Bretthauer J., Anker S. D., Pinet F., et al. (2013). Circulating miR-133a and miR-423-5p fail as biomarkers for left ventricular remodeling after myocardial infarction. *Int. J. Cardiol.* 168, 1837–1840. 10.1016/j.ijcard.2012.12.074

Bayés-Genis A., Lanfear D. E., de Ronde M. W. J., Lupón J., Leenders J. J., Liu Z., et al. (2018). Prognostic value of circulating microRNAs on heart failure-related morbidity and mortality in two large diverse cohorts of general heart failure patients. *Eur. J. Heart Fail.* 20, 67–75.

Beaumont J., López B., Ravassa S., Hermida N., José G. S., Gallego I., et al. (2017). MicroRNA-19b is a potential biomarker of increased myocardial collagen cross-linking in patients with aortic stenosis and heart failure. *Sci. Rep.* 7, 40696.

Beekman M., Nederstigt C., Suchiman H. E. D., Kremer D., van der Breggen R., Lakenberg N., et al. (2010). Genome-wide association study (GWAS)-identified disease risk alleles do not compromise human longevity. *Proc. Natl. Acad. Sci. U. S. A* 107, 18046–18049.

Beg F., Wang R., Saeed Z., Devaraj S., Masoor K., Nakshatri H. (2017). Inflammation-associated microRNA changes in circulating exosomes of heart failure patients. *BMC Res. Notes* 10, 751.

Bekkering S., van den Munckhof I., Nielen T., Lamfers E., Dinarello C., Rutten J., et al. (2016). Innate immune cell activation and epigenetic remodeling in symptomatic and asymptomatic atherosclerosis in humans *in vivo*. *Atherosclerosis* 254, 228–236.

Białek S., Górko D., Zajkowska A., Kołtowski Ł., Grabowski M., Stachurska A., et al. (2015). Release kinetics of circulating miRNA-208a in the early phase of myocardial infarction. *Kardiologia Polska* 73, 613–619.

Bildirici A. E., Arslan S., Özbilüm Şahin N., Berkan Ö., Beton O., Yilmaz M. B. (2018). MicroRNA-221/222 expression in atherosclerotic coronary artery plaque versus internal mammarian artery and in peripheral blood samples. *Biomarkers* 23, 670–675.

BLUEPRINT Consortium (2016). Quantitative comparison of DNA methylation assays for biomarker development and clinical applications. *Nat. Biotechnol.* 34, 726–737.

Bogdarina I., Welham S., King P. J., Burns S. P., Clark A. J. L. (2007). Epigenetic modification of the renin–angiotensin system in the fetal programming of hypertension. *Circ. Res.* 100, 520–526.

Bye A., Røsjø H., Nauman J., Silva G. J. J., Follestad T., Omland T., et al. (2016). Circulating microRNAs predict future fatal myocardial infarction in healthy individuals—the HUNT study. *J. Mol. Cell Cardiol.* 97, 162–168.

Cakmak H. A., Coskunpinar E., Ikitimur B., Barman H. A., Karadag B., Tiryakioglu N. O., et al. (2015). The prognostic value of circulating microRNAs in heart failure: preliminary results from a genome-wide expression study. *J. Cardiovasc. Med. (Hagerstown)* 16, 431–437.

Cao J., Yan Q. (2012). Histone ubiquitination and deubiquitination in transcription, DNA damage response, and cancer. *Front. Oncol.* 2, 26.

Charrier H., Cuvellez M., Dubois-Deruy E., Mulder P., Richard V., Bauters C., et al. (2019). Integrative system biology analyses identify seven microRNAs to predict heart failure. *Noncoding RNA* 5, E22–E30.

Chelbi S. T., Mondon F., Jammes H., Buffat C., Mignot T.-M., Tost J., et al. (2007). Expressional and epigenetic alterations of placental serine protease inhibitors: SERPINA3 is a potential marker of preeclampsia. *Hypertension* 49, 76–83.

Chen F., Yang J., Li Y., Wang H. (2018. a). Circulating microRNAs as novel biomarkers for heart failure. *Hellenic J. Cardiol.* 59, 209–214.

Chen J., Xu L., Hu Q., Yang S., Zhang B., Jiang H. (2015. a). MiR-17-5p as circulating biomarkers for the severity of coronary atherosclerosis in coronary artery disease. *Int. J. Cardiol.* 197, 123–124.

Chen M.-C., Chang T.-H., Chang J.-P., Huang H.-D., Ho W.-C., Lin Y.-S., et al. (2016). Circulating miR-148b-3p and miR-409-3p as biomarkers for heart failure in patients with mitral regurgitation. *Int. J. Cardiol.* 222, 148–154.

Chen S., Chen R., Zhang T., Lin S., Chen Z., Zhao B., et al. (2018. b). Relationship of cardiovascular disease risk factors and noncoding RNAs with hypertension: a case-control study. *BMC Cardiovasc. Disord.* 18, 58.

Chen W., Li S. (2017). Circulating microRNA as a novel biomarker for pulmonary arterial hypertension due to congenital heart disease. *Pediatr. Cardiol.* 38, 86–94.

Chen X., Ba Y., Ma L., Cai X., Yin Y., Wang K., et al. (2008). Characterization of microRNAs in serum: a novel class of biomarkers for diagnosis of cancer and other diseases. *Cell Res.* 18, 997–1006.

Chen X., Zhang L., Su T., Li H., Huang Q., Wu D., et al. (2015. b). Kinetics of plasma microRNA-499 expression in acute myocardial infarction. *J. Thorac. Dis.* 7, 890–896.

Cheng C., Wang Q., You W., Chen M., Xia J. (2014). MiRNAs as biomarkers of myocardial infarction: a meta-analysis. *PLoS ONE* 9, e88566.

Cho H.-M., Lee H.-A., Kim H. Y., Han H. S., Kim I. K. (2011). Expression of Na⁺-K⁺ - 2Cl⁻ cotransporter 1 is epigenetically regulated during postnatal development of hypertension. *Am. J. Hypertens.* 24, 1286–1293.

Choi J.-H., Nam K.-H., Kim J., Baek M. W., Park J.-E., Park H.-Y., et al. (2005). Trichostatin A exacerbates atherosclerosis in low density lipoprotein receptor-deficient mice. *Arteriosclerosis, Thrombosis, and Vascular Biol.* 25, 2404–2409.

Choi S. Y., Yun J., Lee O. J., Han H. S., Yeo M. K., Lee M. A., et al. (2013). MicroRNA expression profiles in placenta with severe preeclampsia using a PNA-based microarray. *Placenta* 34, 799–804.

Cortez-Dias N., Costa M. C., Carrilho-Ferreira P., Silva D., Jorge C., Calisto C., et al. (2016). Circulating miR-122-5p/miR-133b ratio is a specific early prognostic biomarker in acute myocardial infarction. *Circ. J.* 80, 2183–2191.

Coskunpinar E., Cakmak H. A., Kalkan A. K., Tiryakioglu N. O., Erturk M., Ongen Z. (2016). Circulating miR-221-3p as a novel marker for early prediction of acute myocardial infarction.

Gillette TG, & Hill JA (2015). Readers, writers, and erasers: chromatin as the whiteboard of heart disease. *Circ Res*, 116, 1245–1253. [PMC free article] [PubMed] [Google Scholar]

Jeltsch A, & Jurkowska RZ (2014). New concepts in DNA methylation. *Trends Biochem Sci*, 39, 310–318. [PubMed] [Google Scholar]

Raghuraman S., Donkin I., Verstehey S., Barrès R., Simar D. The emerging role of epigenetics in inflammation and immunometabolism. *Trends in Endocrinology & Metabolism*. 2016;27(11):782–795.doi:10.1016/j.tem.2016.06.008. [PubMed] [CrossRef] [Google Scholar]

Thom T, Haase N, Rosamond W, Howard VJ, Rumsfeld J, et al. Heart disease and stroke statistics–2006 update: a report from the American Heart Association Statistics Committee and Stroke Statistics Subcommittee. *Circulation*. 2006;113:e85–151. [PubMed] [Google Scholar]

Yang Y, Hsu PJ, Chen YS, & Yang YG (2018). Dynamic transcriptomic m(6)A decoration: writers, erasers, readers and functions in RNA metabolism. *Cell Res*, 28, 616–624. [PMC free article] [PubMed] [Google Scholar]

Yang Y, Li X, Peng L, An L, Sun N, Hu X, Zhou P, Xu Y, Li P, & Chen J (2018). Tanshindiol C inhibits oxidized low-density lipoprotein induced macrophage foam cell formation via a peroxiredoxin 1 dependent pathway. *Biochim Biophys Acta*, 1864, 882–890. [PubMed] [Google Scholar]

